

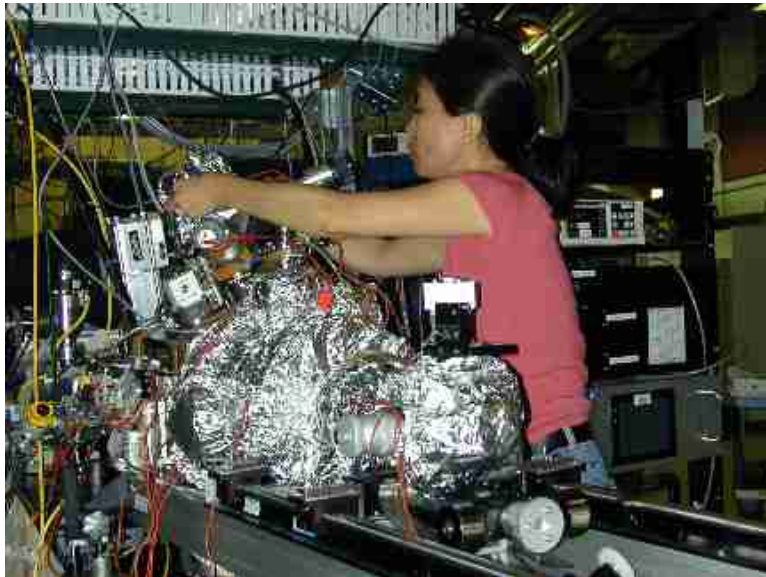
Diffraction microscopy of biological specimens: imaging of a freeze-dried yeast cell

David Shapiro, Tobias Beetz, Chris Jacobsen, Janos Kirz, Enju Lima, Huijie Miao, Aaron Neiman, David Sayre: Stony Brook

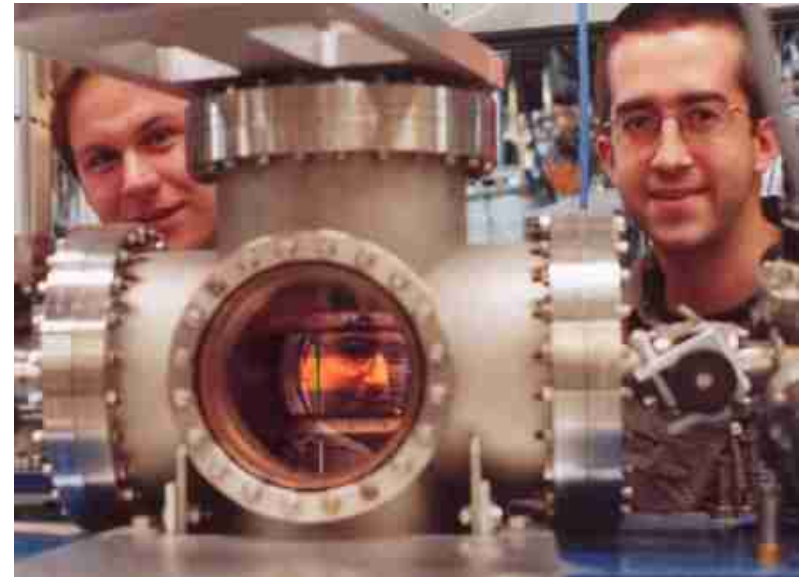
Pierre Thibault, Veit Elser: Cornell

Malcolm Howells: Advanced Light Source

(To be published in Proc. Nat. Acad. Sci.)



Enju Lima



**Tobias
Beetz**

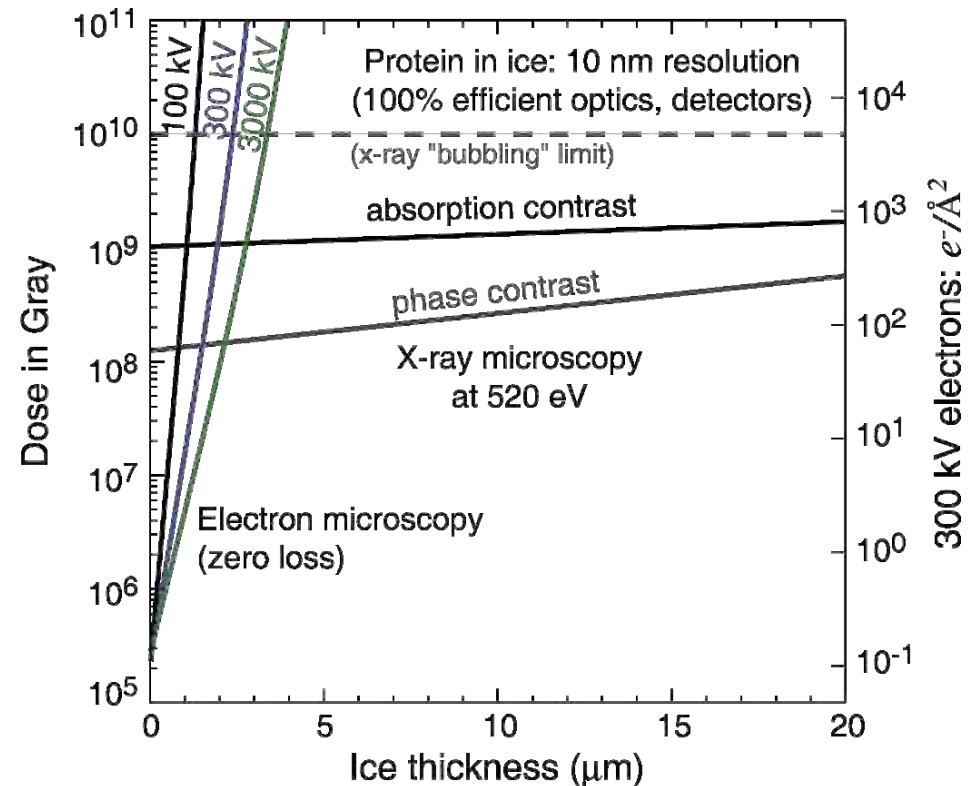
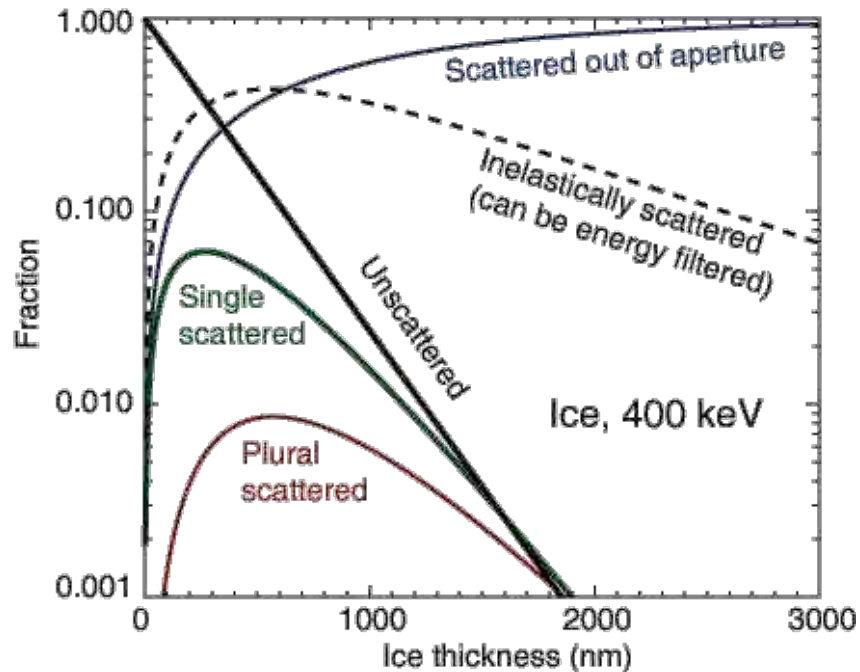
**David
Shapiro**

Synchrotron imaging in context

- Light microscopy:
 - Live cells at ~200 nm resolution. Approaching 50 nm with spatial light modulation! Gustafsson et al., UC San Francisco; Cremer et al., Heidelberg
 - Fluorescent labels for specific proteins (permeating, injected, or genetically "programmed in" such as Green Fluorescent Protein and its variants)
- Electron microscopy:
 - 0.1-0.2 nm resolution on robust, crystalline samples.
 - 0.5-1 nm resolution on many copies of identical molecules (single particle methods)
 - 6 nm resolution in tomography
 - Immunogold labeling

X rays and thick specimens

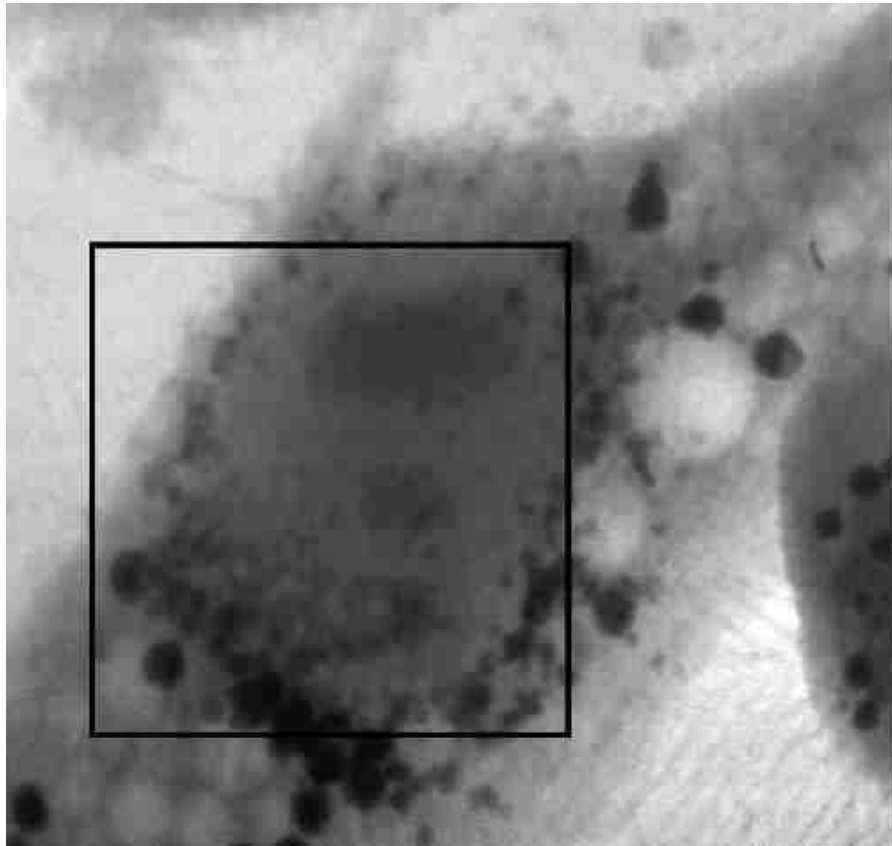
X rays better for thick specimens. Sayre et al., *Science* **196**, 1339 (1977); Schmahl & Rudolph in **X-ray Microscopy: Instrumentation and Biological Applications** (Springer, 1987)



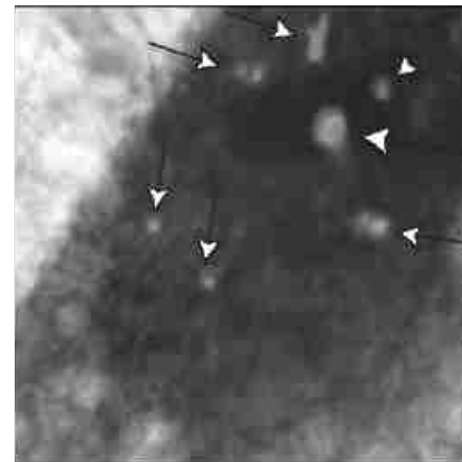
These plots: Jacobsen, Medenwaldt, and Williams, in **X-ray Microscopy & Spectromicroscopy** (Springer, 1998)

Radiation damage resistance of wet specimens at liquid nitrogen temperature

Left: frozen hydrated image **after** exposing several regions to $\sim 10^{10}$ Gray



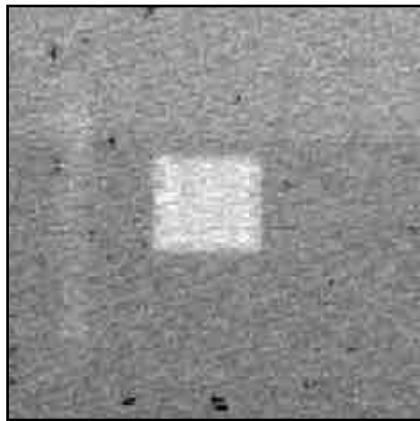
Right: after warmup in microscope (eventually freeze-dried): holes indicate irradiated regions!



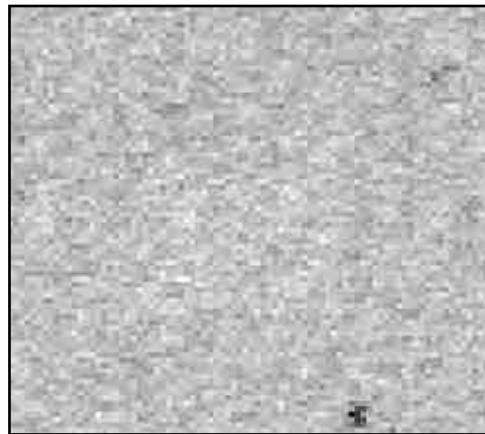
— 7 μm

PMMA at room, LN2 temperature

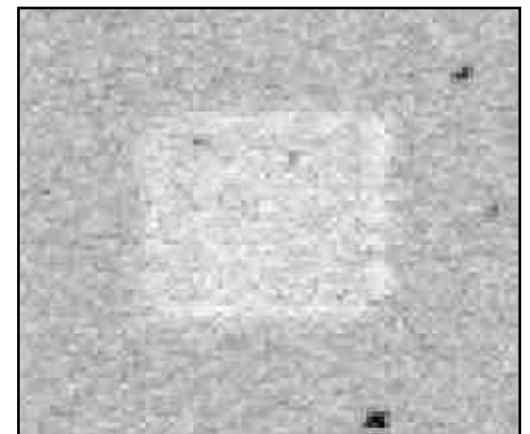
- PMMA: poly methyl methacrylate (plexiglass!) which is especially radiation sensitive - it's used as a resist for electron beam lithography
- Repeated sequence: dose (small square), spectrum (defocused beam).
Low dose, larger area image at end.



Room temperature:
mass loss
immediately visible



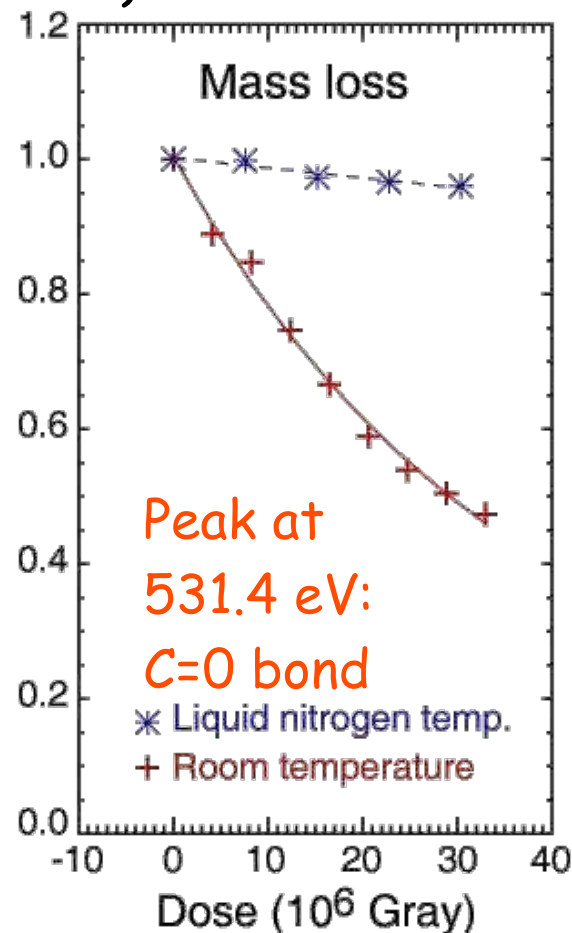
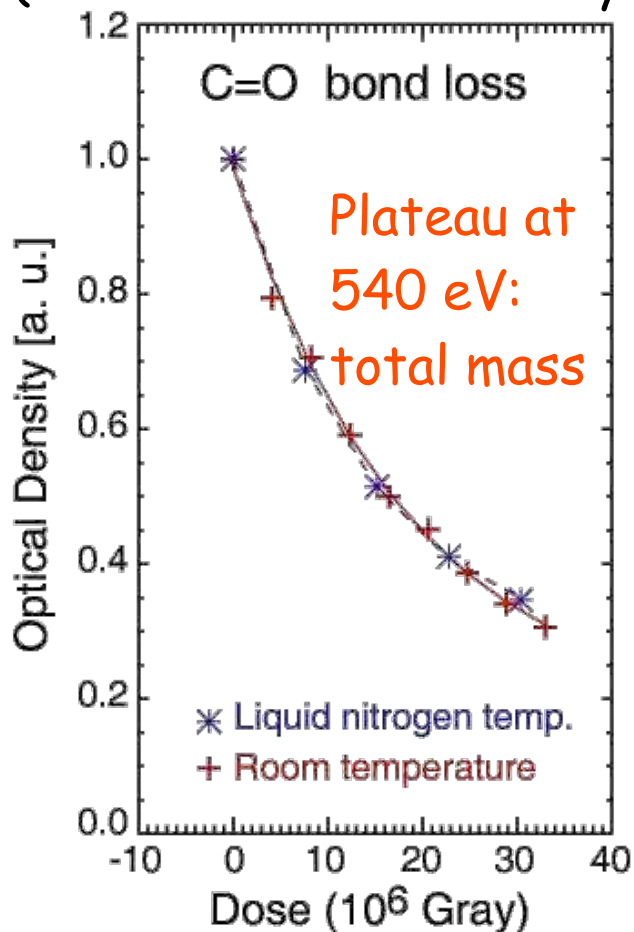
LN2 temperature:
no mass loss
immediately visible



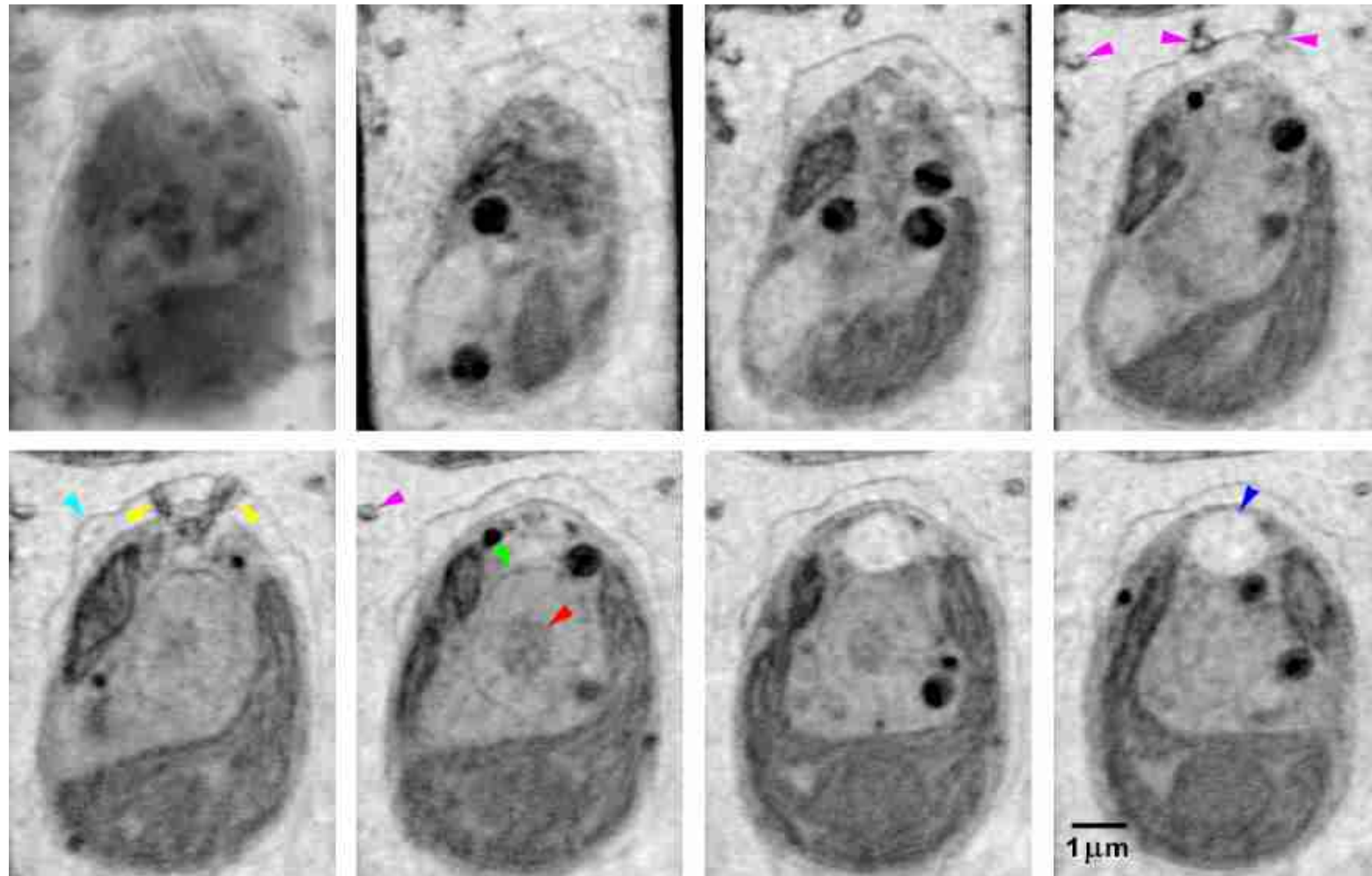
After warm-up:
mass loss becomes
visible

Results from fitting spectra

LN₂ temp: protection against mass loss, but not against breaking bonds (at least C=O bond in dry PMMA)



Soft x-ray tomography of algae shock-frozen in liquid ethane: Slices through a tomographic reconstruction

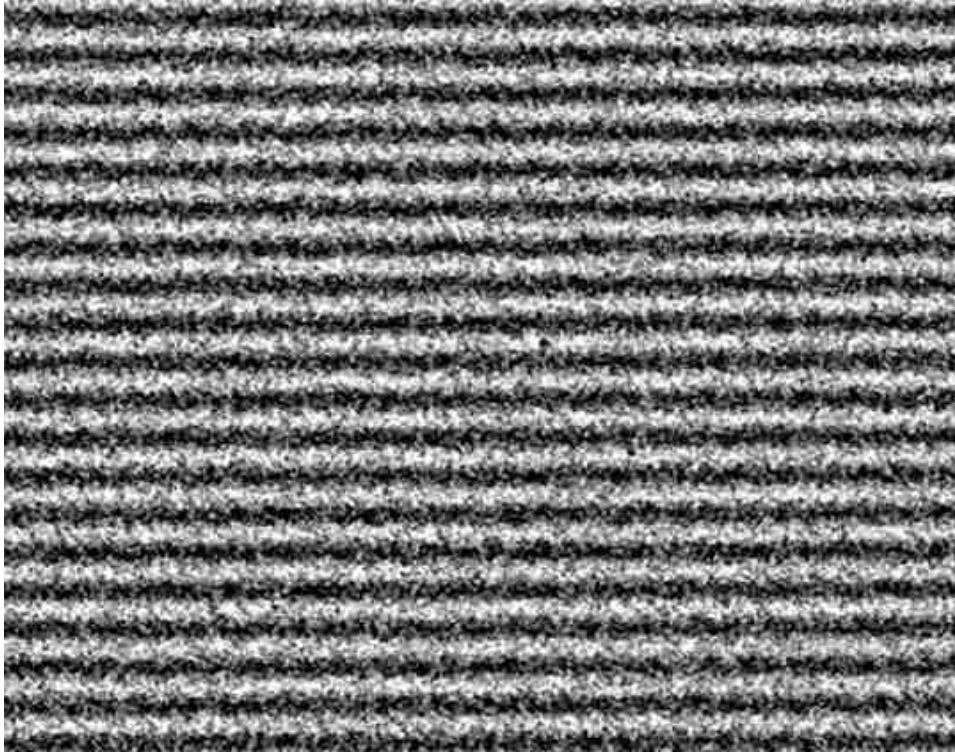


- Vacuole
- Nuclear membrane
- Nucleolus
- Flagellar roots and neuromotor
- Cell wall
- Flagella

Weiss, Schneider et al., *Ultramicroscopy* **84**, 185 (2000).

See also Larabell and Le Gros, *Molecular Biology of the Cell* **115**, 957 (2004)

Resolution frontier of zone plates



But efficiency only ~3%.
Other results: 9.2% at 20
nm: Peuker, Appl. Phys. Lett.
78, 2208 (2001)

15.1 nm half-pitch multilayer slice imaged
with a 15 nm outermost zone width zone
plate. Chao et al., CXRO/LBL. Previous work:
see Chao et al., Opt. Lett. **28**, 2019 (2003)

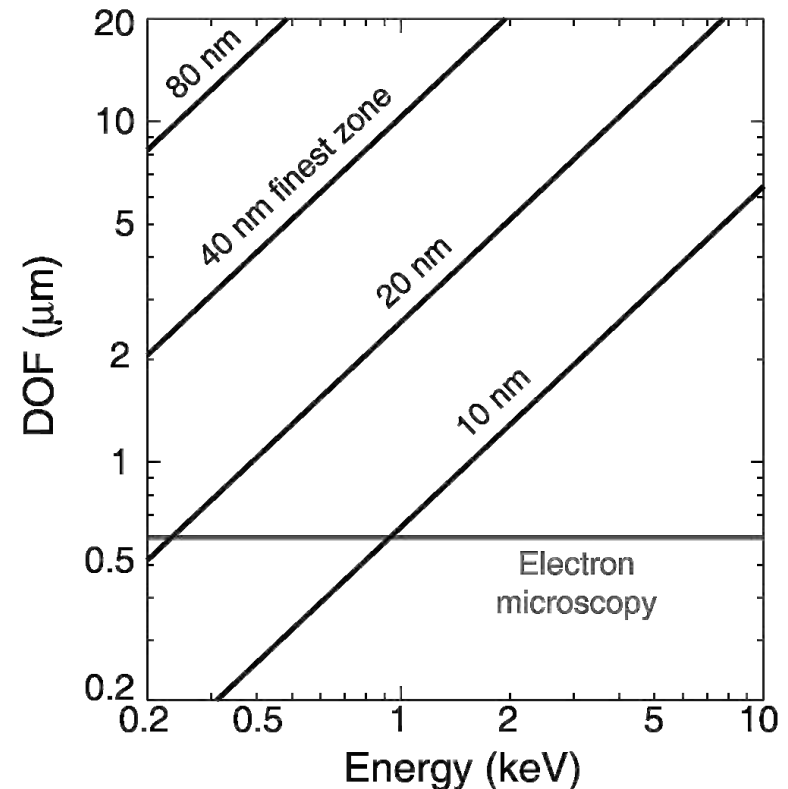
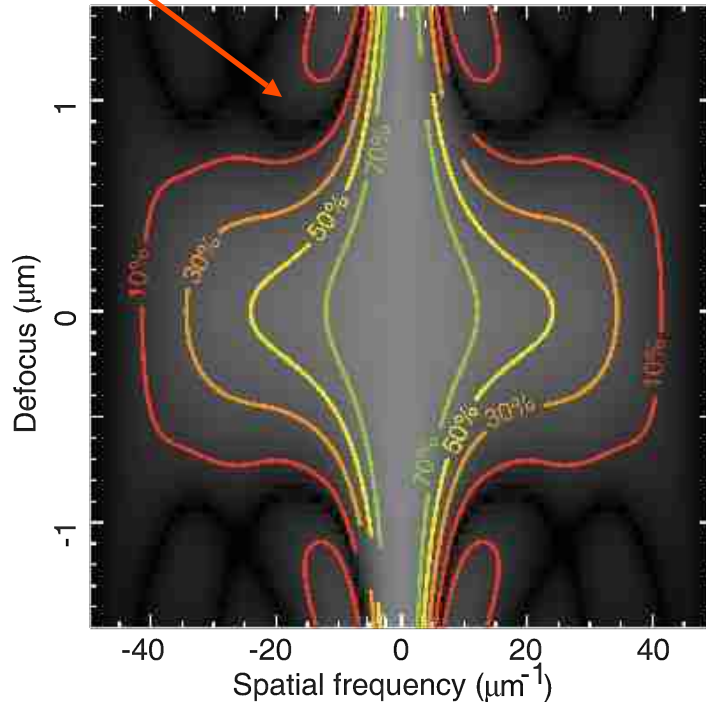
Depth of field limit to conventional tomography

Transverse: $\delta_t \Rightarrow \frac{\lambda}{4\theta} = \frac{dr_N}{2}$

Longitudinal: $\delta_l \Rightarrow \frac{\lambda}{\theta^2} = 8dr_N \frac{dr_N}{\lambda}$

Phase inversion

Contrast versus defocus:
 $\delta_{rN}=20 \text{ nm}, \lambda=2.5 \text{ nm}$

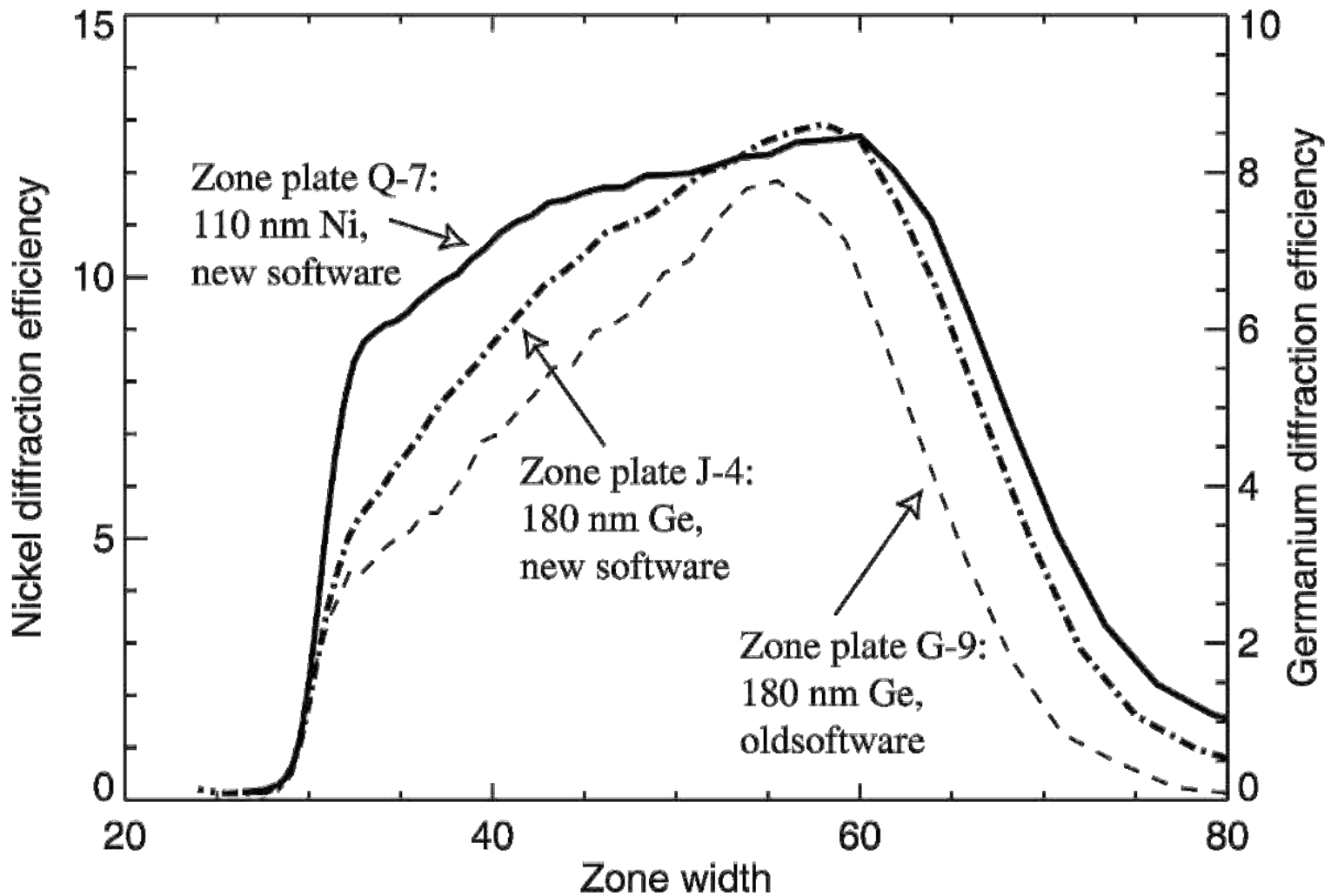


MTF of a 20 nm zone plate @ 0.5 keV

Decreased monochromaticity increases DOF but also reduces transfer function. Schneider et al., Surf. Rev. Lett. **9**, 177 (2002)

Through-focus deconvolution on a complex object requires exact knowledge of partially coherent transfer function

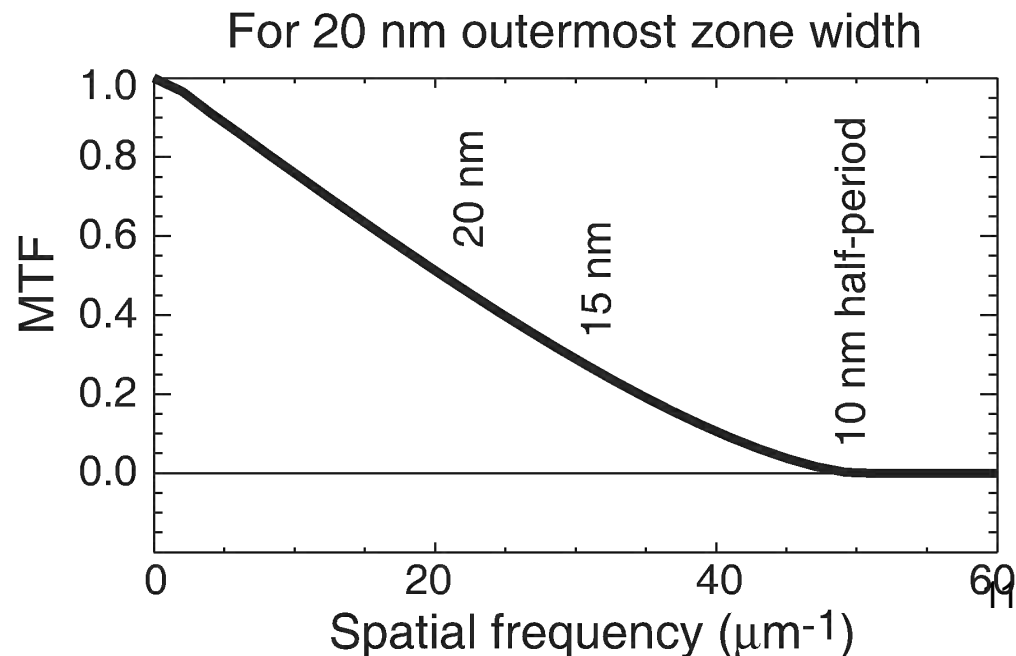
Limitations of zone plates



Radiation damage sets the ultimate resolution limit

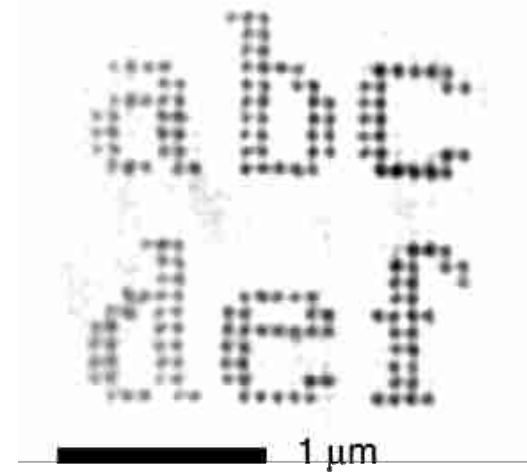
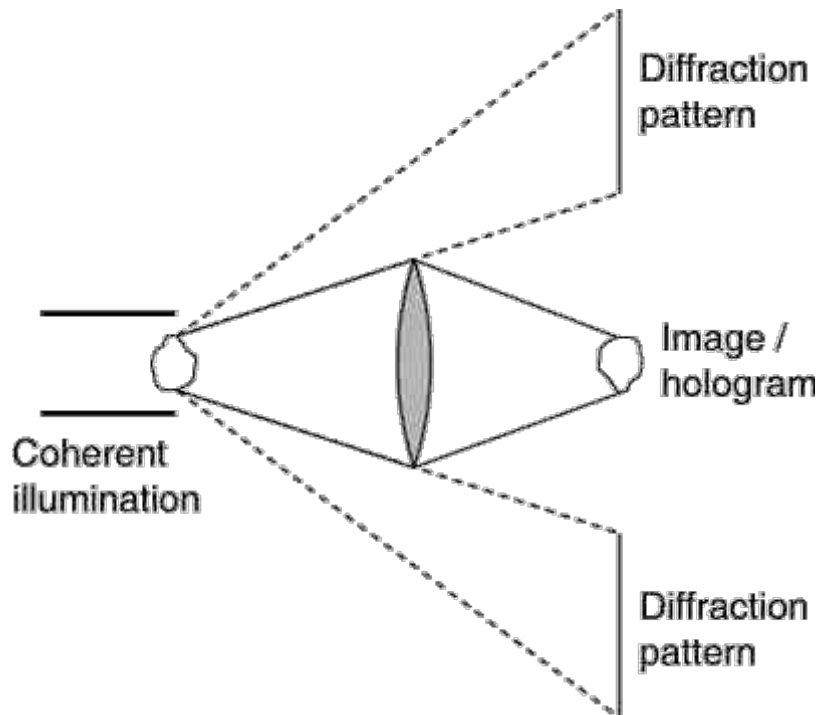
- For many specimens, radiation damage sets the ultimate limit on achievable resolution
- With 20 nm zone plates, have 8x loss for diffraction efficiency, 2x loss for window transmission, 5x loss for modulation transfer function (MTF) at 15 nm feature size.
 - On-Bragg (tilted) zone plates will help: Maser, Opt. Comm. **89**, 355 (1992); Hambach and Schneider, J. Vac. Sci. Tech. B **17**, 3212 (1999).
- Can we avoid this 80x signal loss, and go beyond the limits of available optics?

(MTF=modulation transfer function)



Can we beat the losses of lenses?

- Proposed by Sayre (in Schlenker, ed., *Imaging and Coherence Properties in Physics*, Springer-Verlag, 1980)
- Previous experiments by Sayre, Kirz, Yun, Chapman, Miao



First x-ray
reconstruction: Miao et
al., *Nature* **400**, 342
(1999)¹²

Phase matters

Image \rightarrow Fourier transform \rightarrow zero magnitude or phase \rightarrow inverse Fourier transform



Malcolm Howells
at La Clusaz

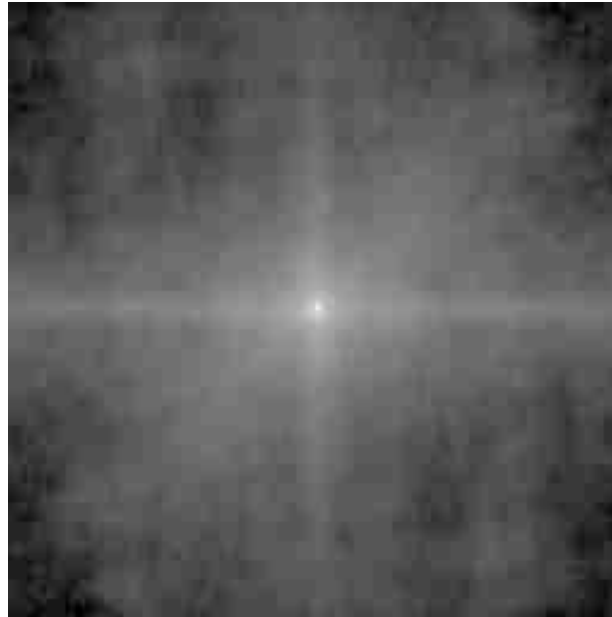


Image using only
Fourier magnitudes



Image using only
Fourier phases

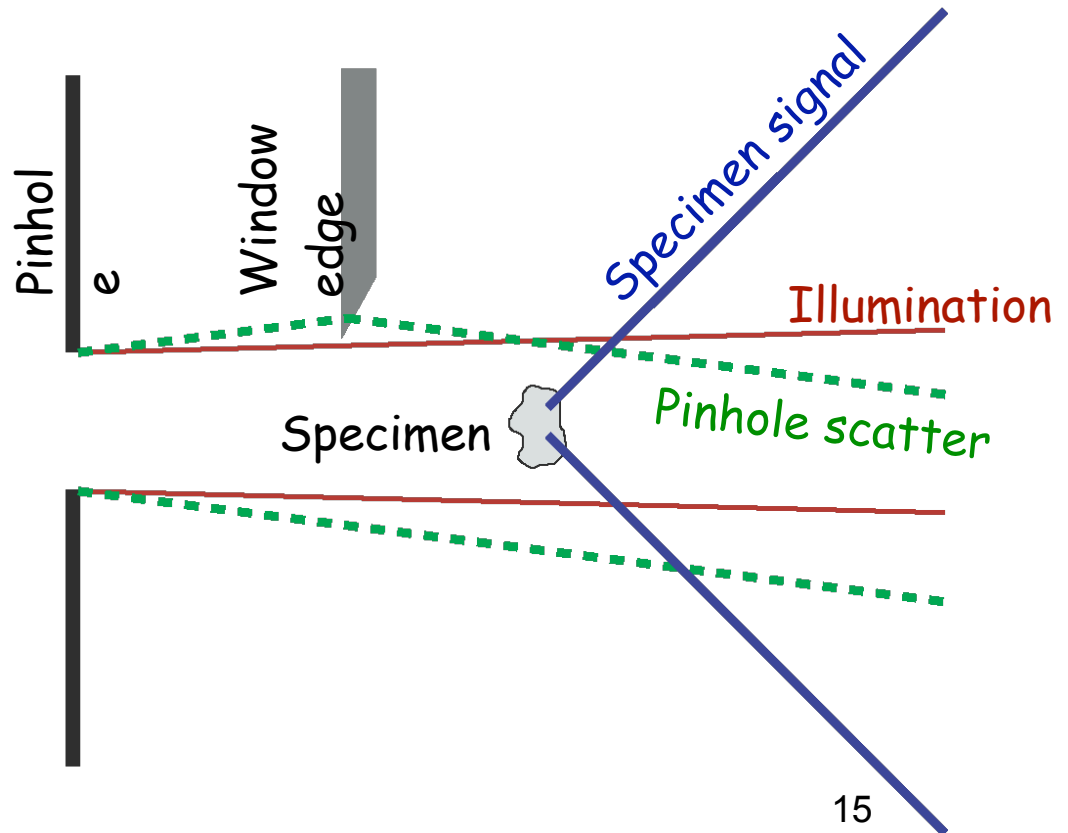
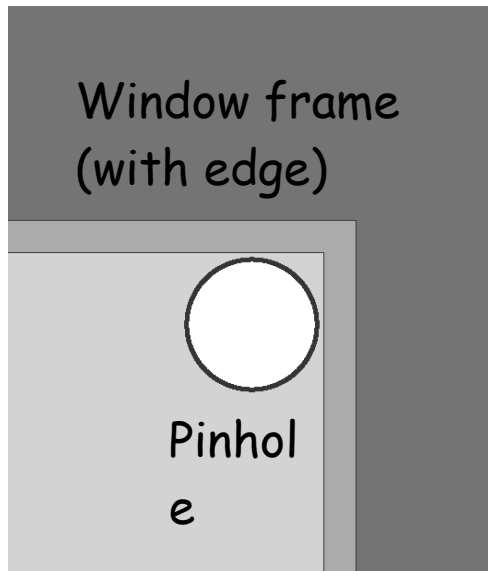
Wrapping our tentacles around the problem



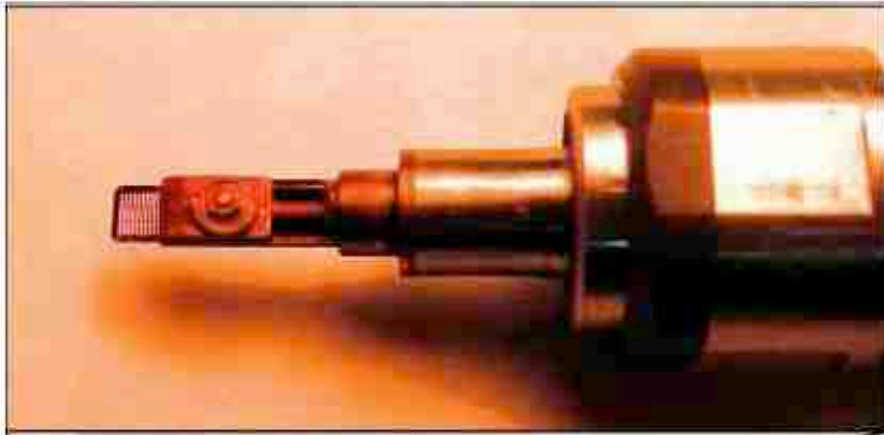
(Porquerolles)

Removing scatter from pinholes

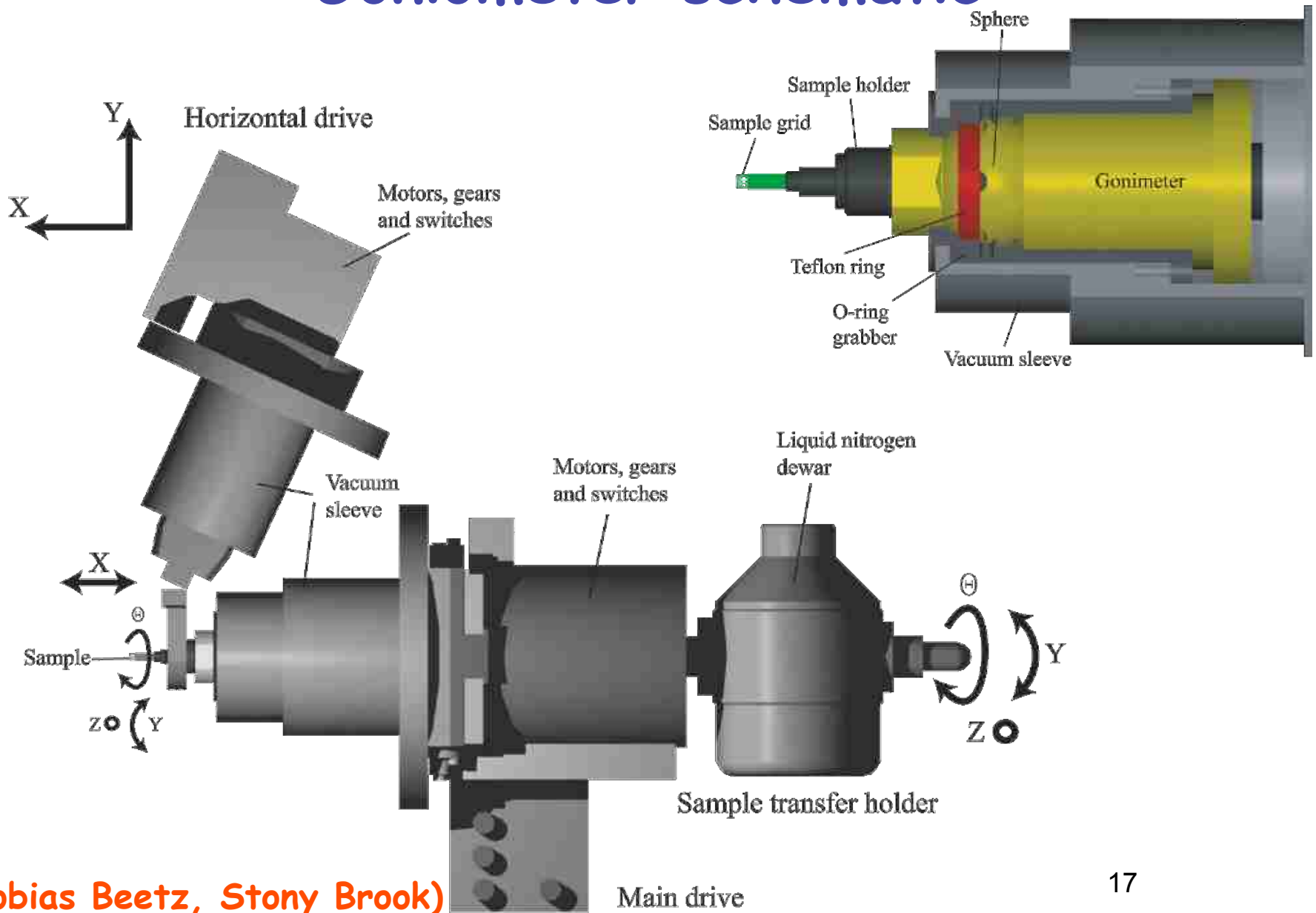
- Pinholes have scatter; can overwhelm weak diffraction.
- Use a "soft," refractive corner to limit to one quadrant (idea due to H. Chapman, then at Stony Brook)



Gatan 630 cryo holder



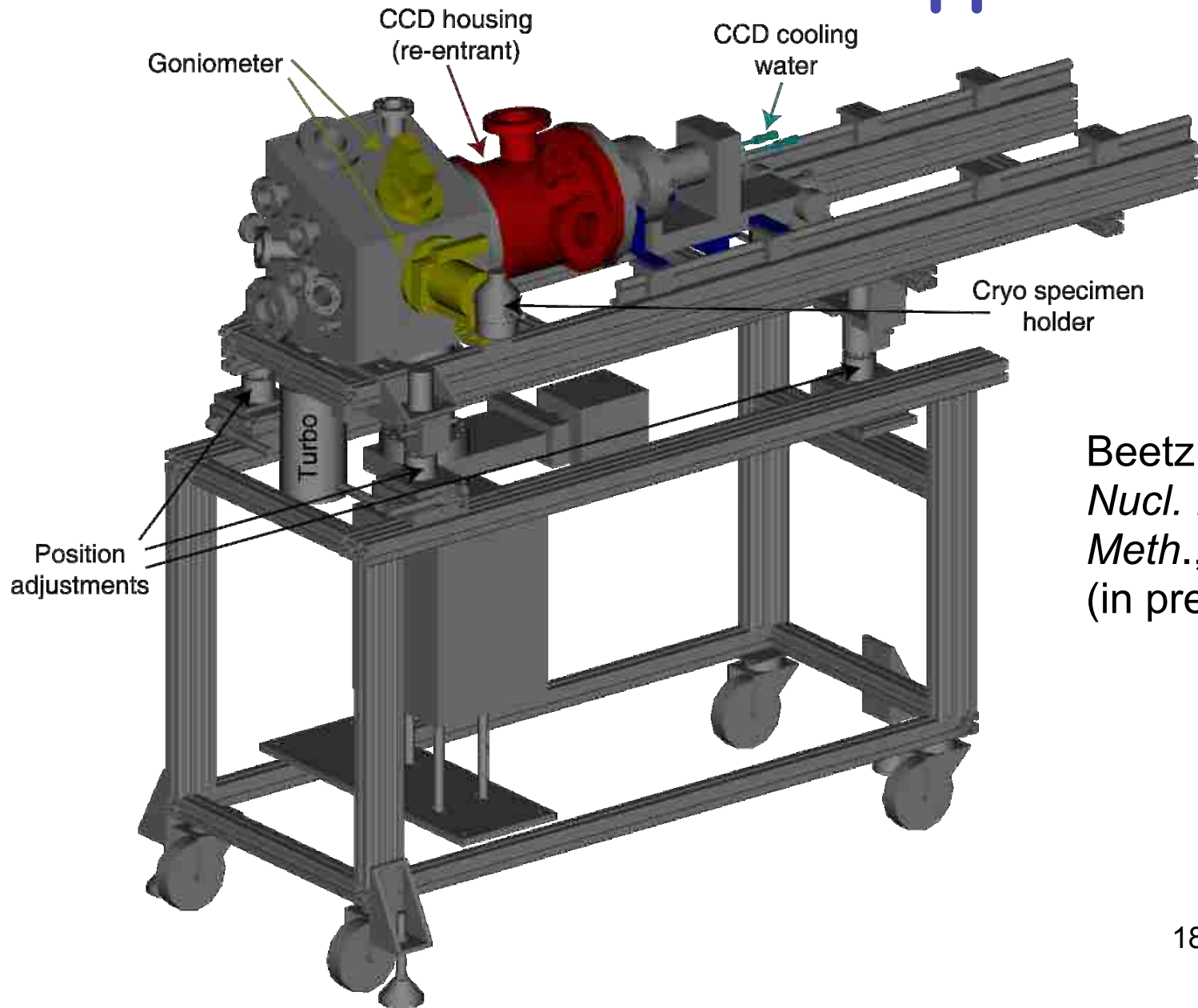
Goniometer schematic



(Tobias Beetz, Stony Brook)

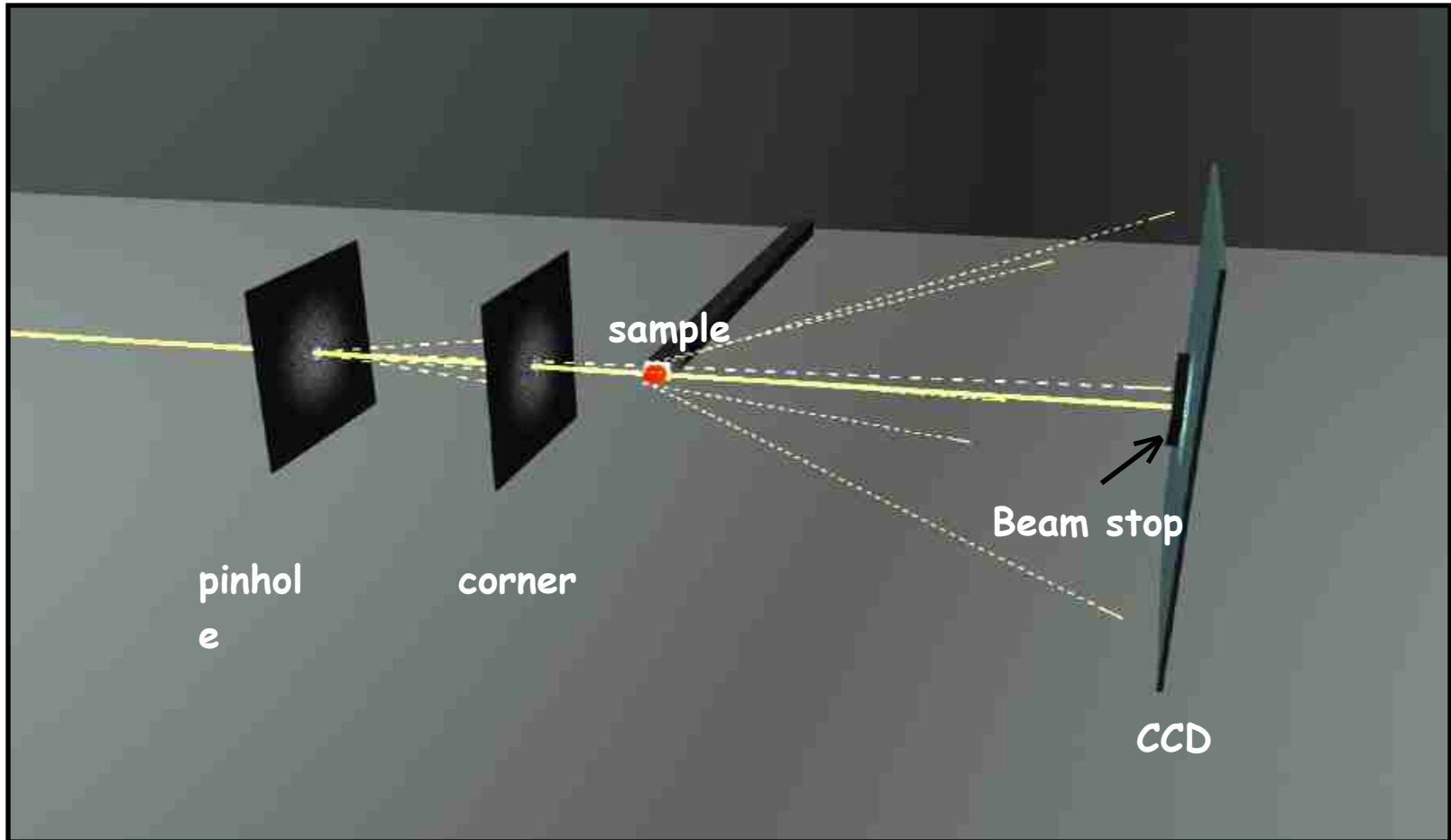
Main drive

ALS beamline 9.0.1 apparatus



Beetz *et al.*,
Nucl. Inst. Meth., 2005
(in press)

Experimental setup

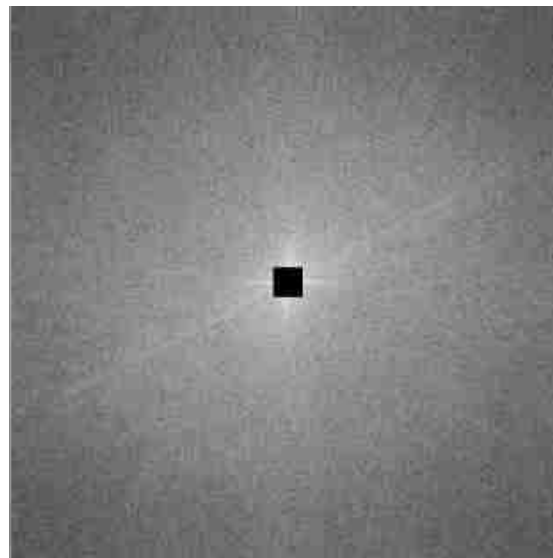


Inside vacuum chamber

(Enju Lima, Stony Brook)

Effect of missing low spatial frequencies

Meghan Sumner



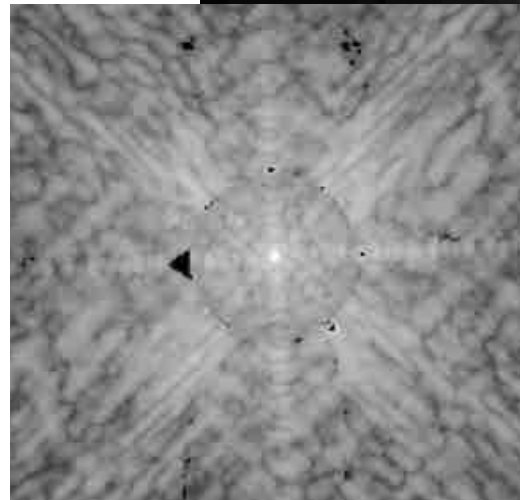
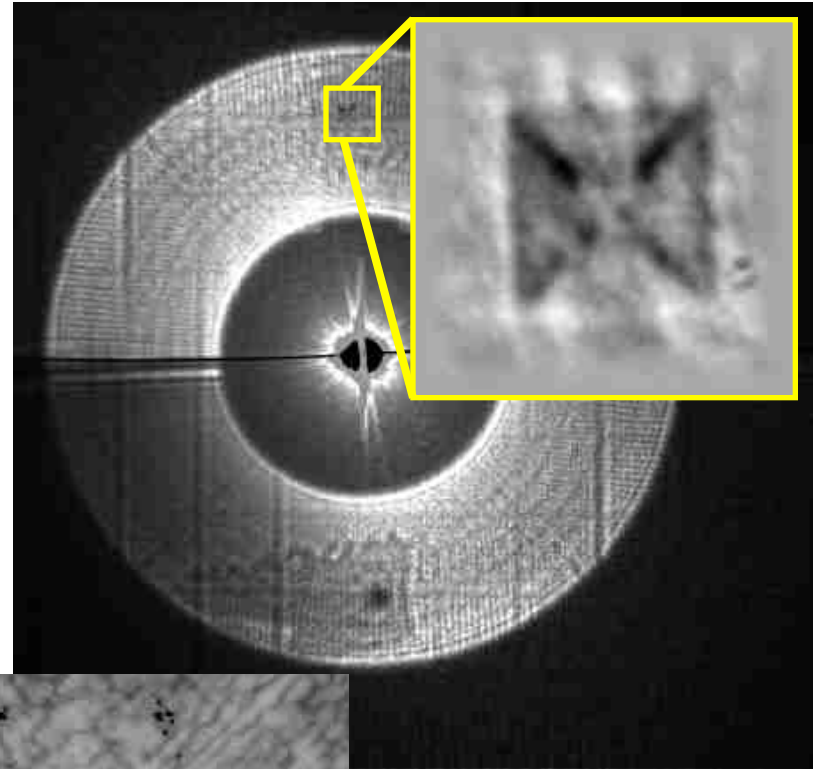
Zone plate imaging

Zone plate can be inserted behind specimen by motor control.

However:

- 20 nm zone plate with central stop, JBX-6000 stitching errors (S. Spector)
- At 16 cm CCD distance; optical magnification gives only 100 nm pixels

Image of H. Chapman's pyramid by T. Beetz



Patch in low spatial frequencies with zone plate image? Counterproductive thus far!
21

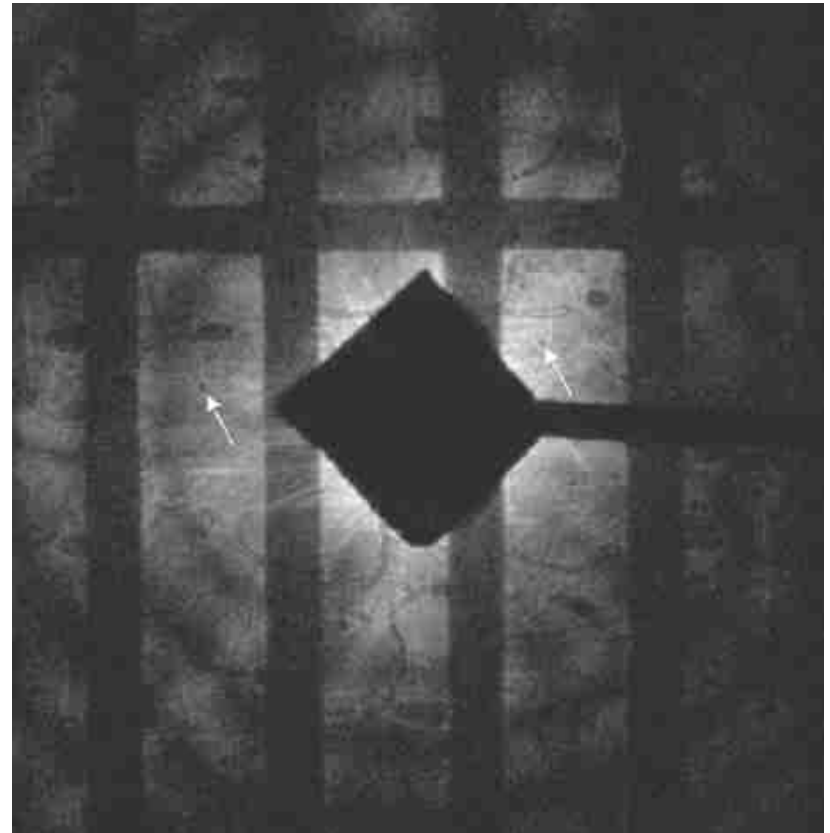
When hopes are dashed on the rocks, you
have to find a way out



(Porquerolles)

Finding specimens

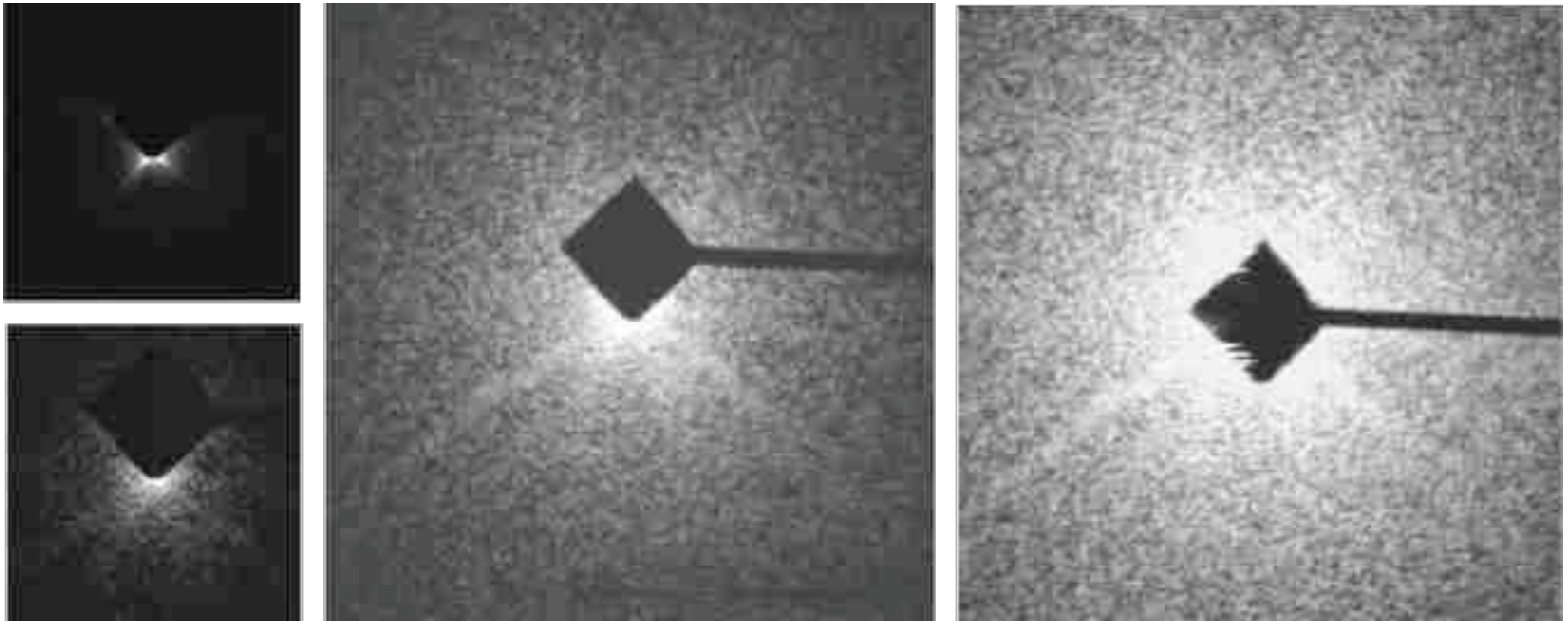
- “Point projection” microscope from moving coherence pinhole upstream.
- Not a good way to evaluate preparations; poor feedback loop!
- Future solution: build a crude scanning microscope into apparatus



Yeast cells on a grid

Dealing with the CCD limits

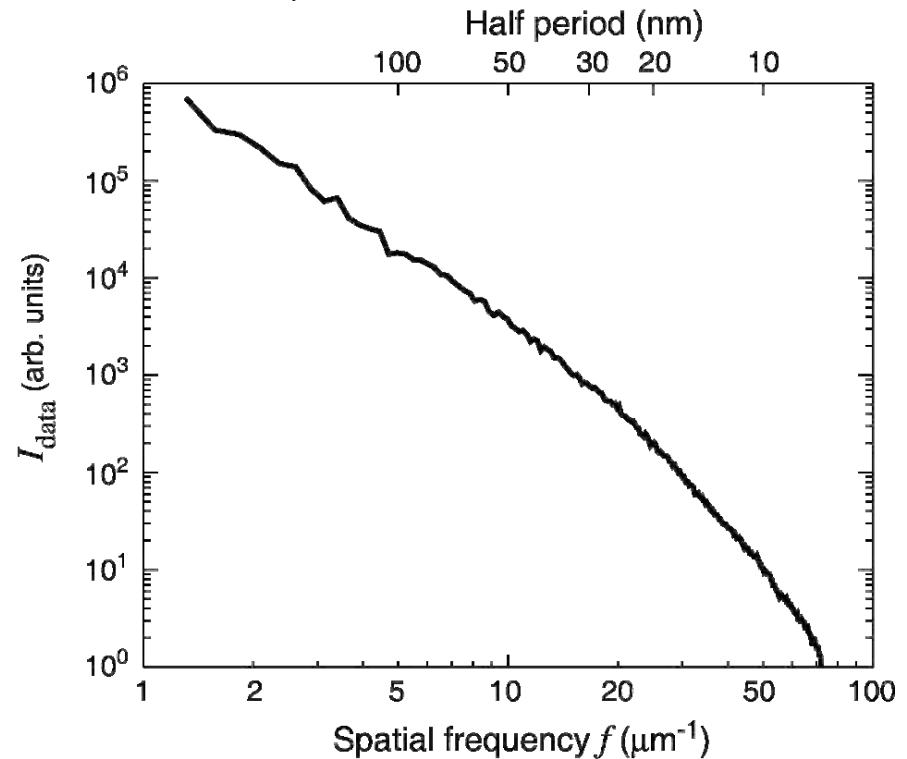
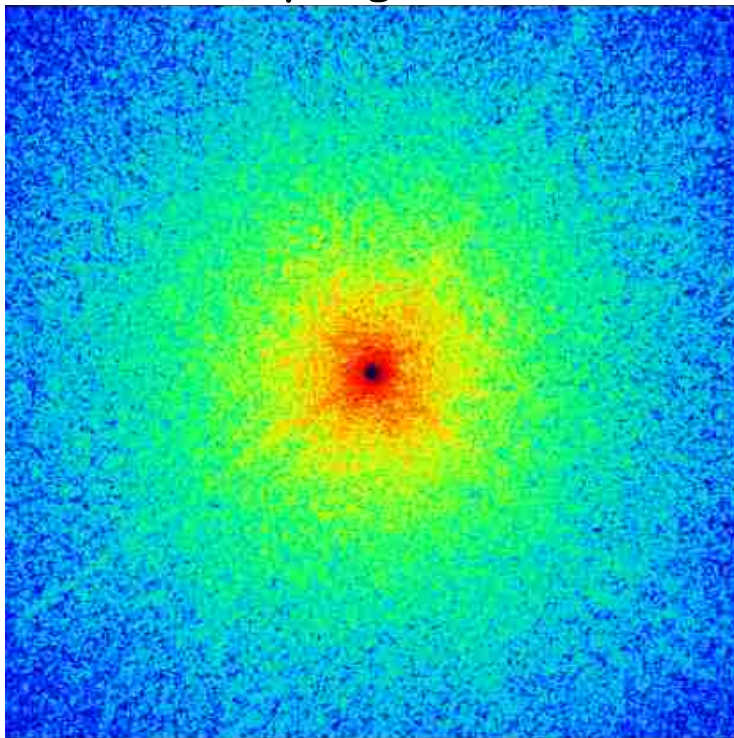
- ~Direct detection: 100 e⁻/photon; ~10⁵ e⁻ full well capacity; ~10³ dynamic range.
- Diffraction data drops off $\propto f^{-4}$
- Move beamstop and merge exposures



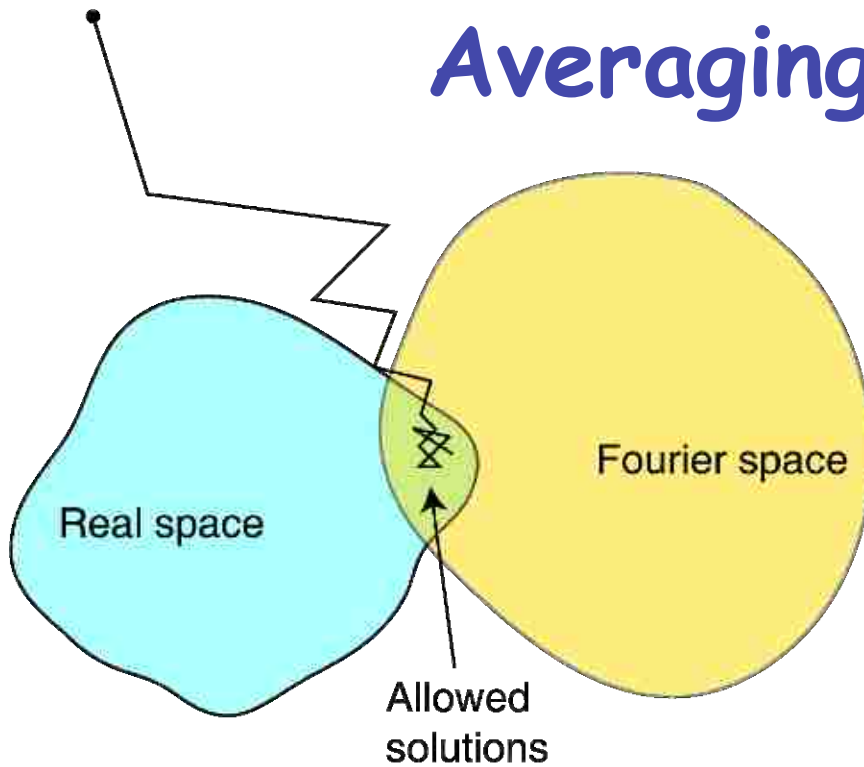
Future: use a "Tower of Hanoi" beamstop?

Diffraction from a yeast cell

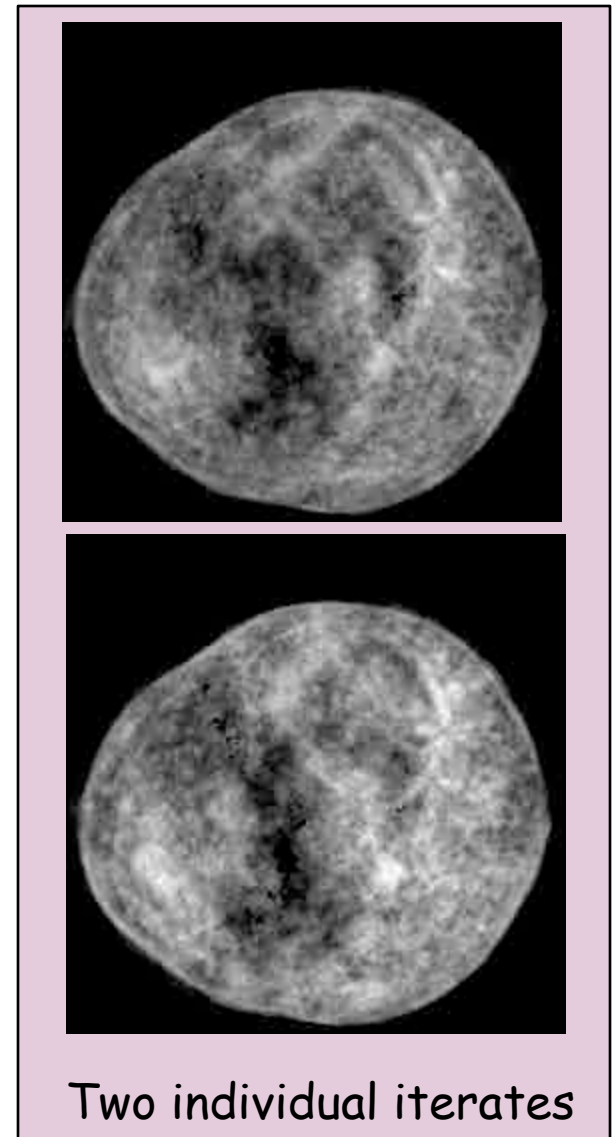
- ALS beamline 9.0.1 operated at 750 eV
- Total dose to freeze-dried, room temperature cell around 10^8 Gray
- Oversampling ratio is about 9 in each dimension



Averaging iterates



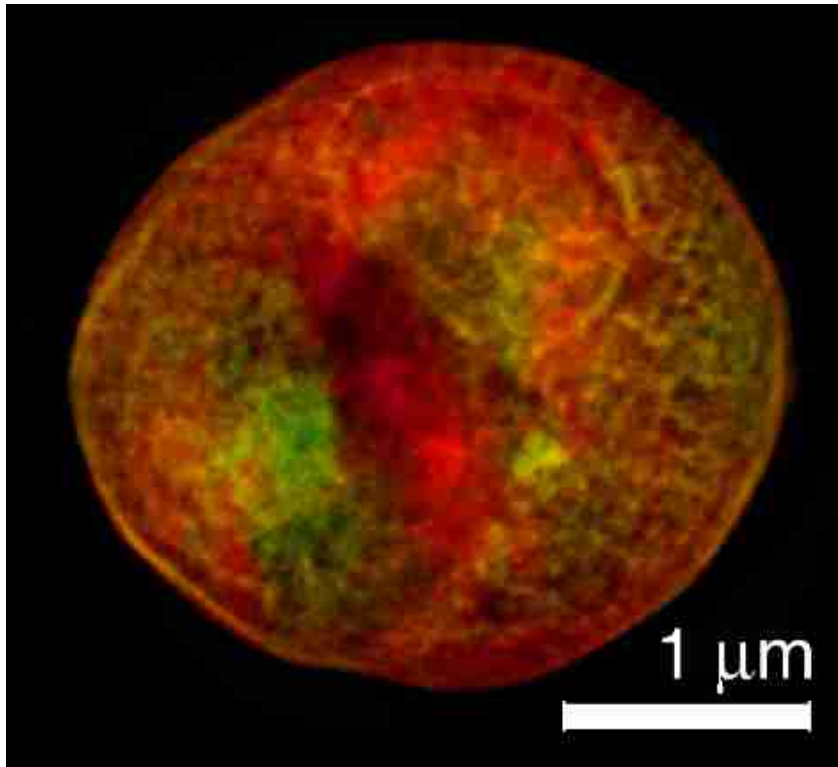
- Ideally, there is a single point of best agreement between various constraints.
- Noise and incomplete constraints only provide a neighborhood of acceptable solutions. Choosing one iterate is unwarranted.
- Averaging many iterates:
 - Reproducible information is reinforced
 - Non-reproducible information is suppressed
- Elser and Thibault, Cornell



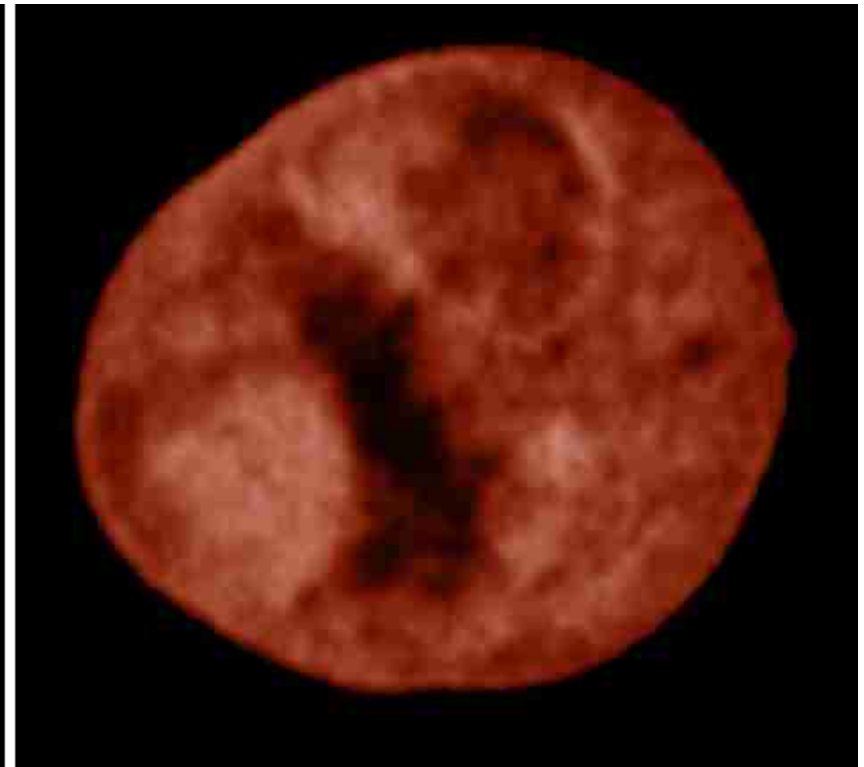
Reconstruction

- Done first by Thibault and Elser; then by Shapiro, Lima
- Difference map algorithm
- Hand-drawn tight support based on autocorrelation and reconstruction attempts
- Shrink-wrap was not used because it started to “eat into” the cell but this could probably be improved.

Comparison with a microscope



Diffraction reconstruction
(data taken at 750 eV;
absorption as brightness,
phase as hue).



Stony Brook/NSLS STXM
image with 45 nm Rayleigh
resolution zone plate at 520
eV (absorption as
brightness)

Using color to represent phase



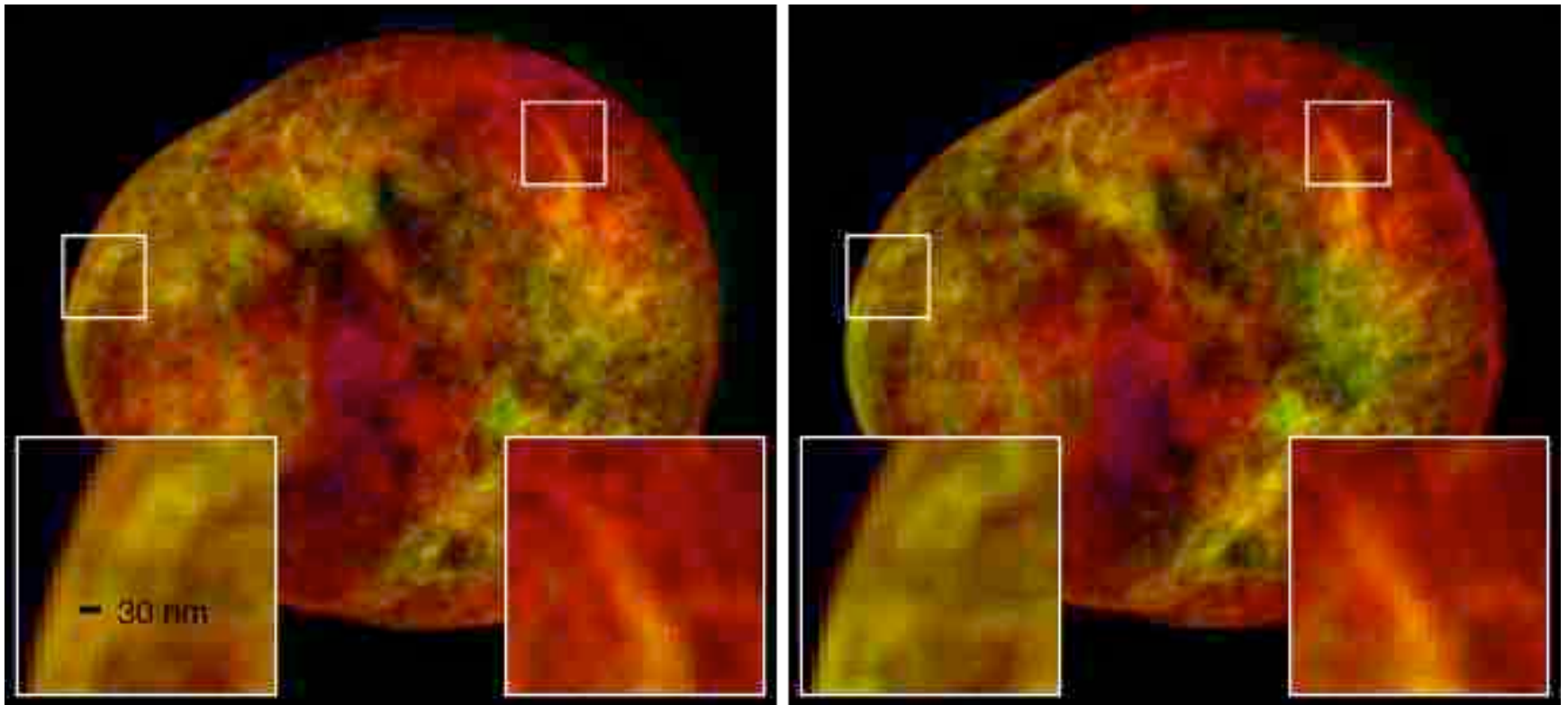
(Porquerolles)

Are we only seeing things we want to see?



(HMS Rhone, British Virgin Islands)

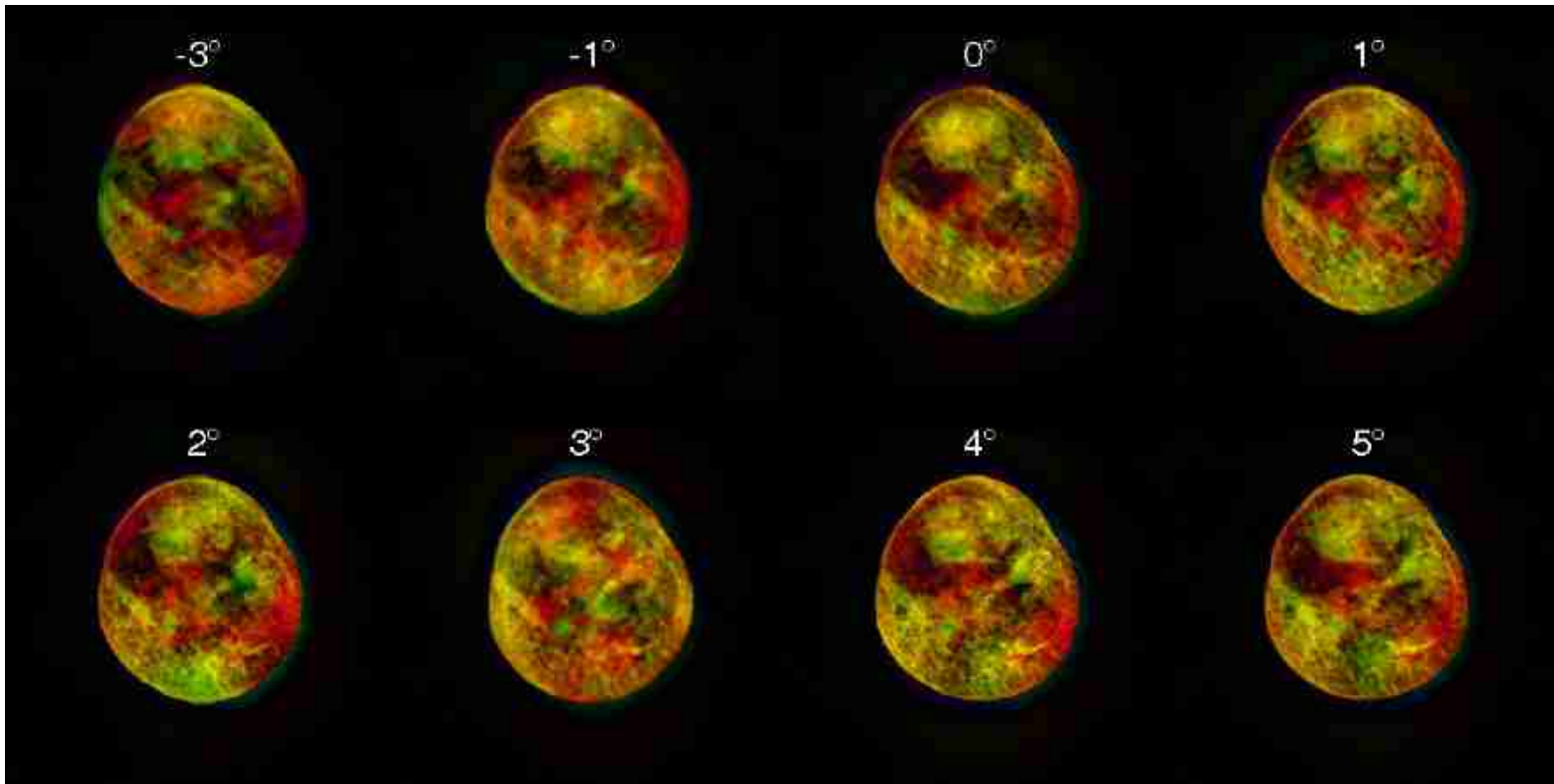
Data taken 1° apart



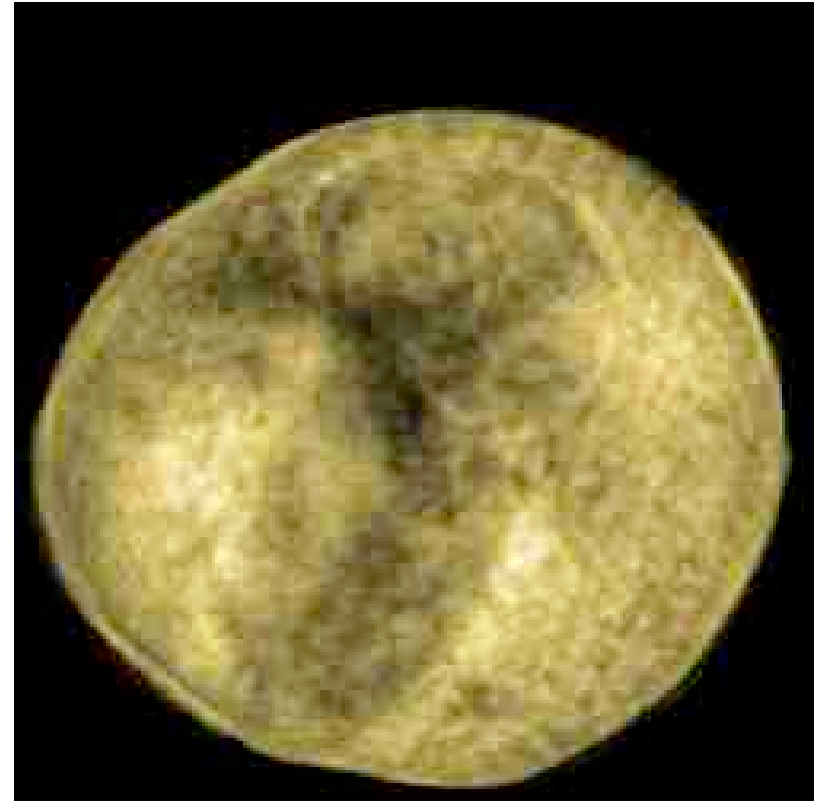
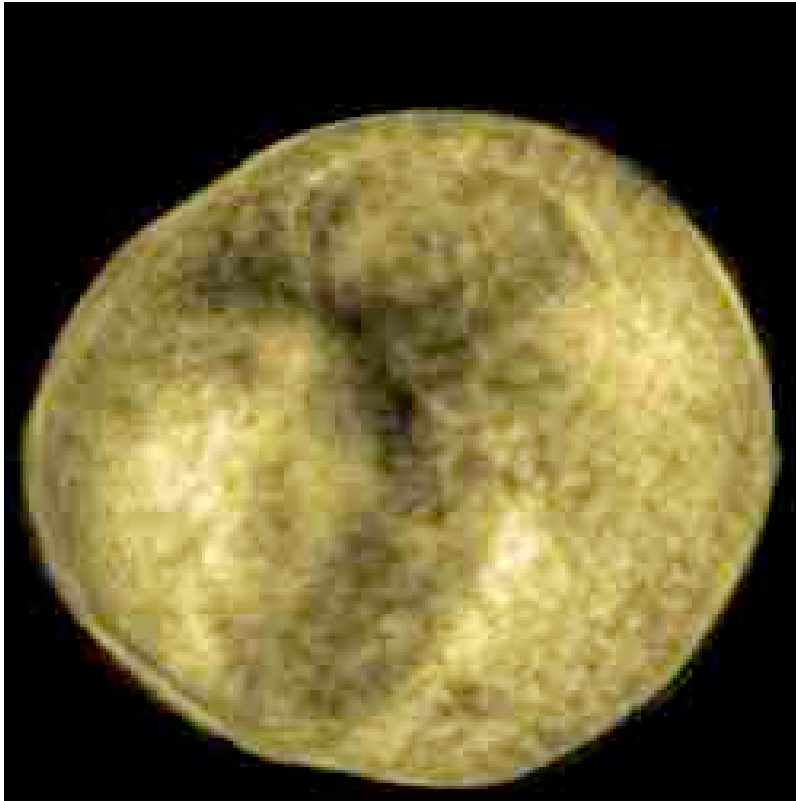
Separate reconstructions of separate data sets give reconstructions that are quite consistent

Tilt series

This sample: only an 8° tilt range. 2D reconstructions on each projection.



Different starting random phases



Two separate runs of algorithm with different random starting phases. In both cases, 125 iterates spaced 40 iterations apart were averaged (E. Lima).

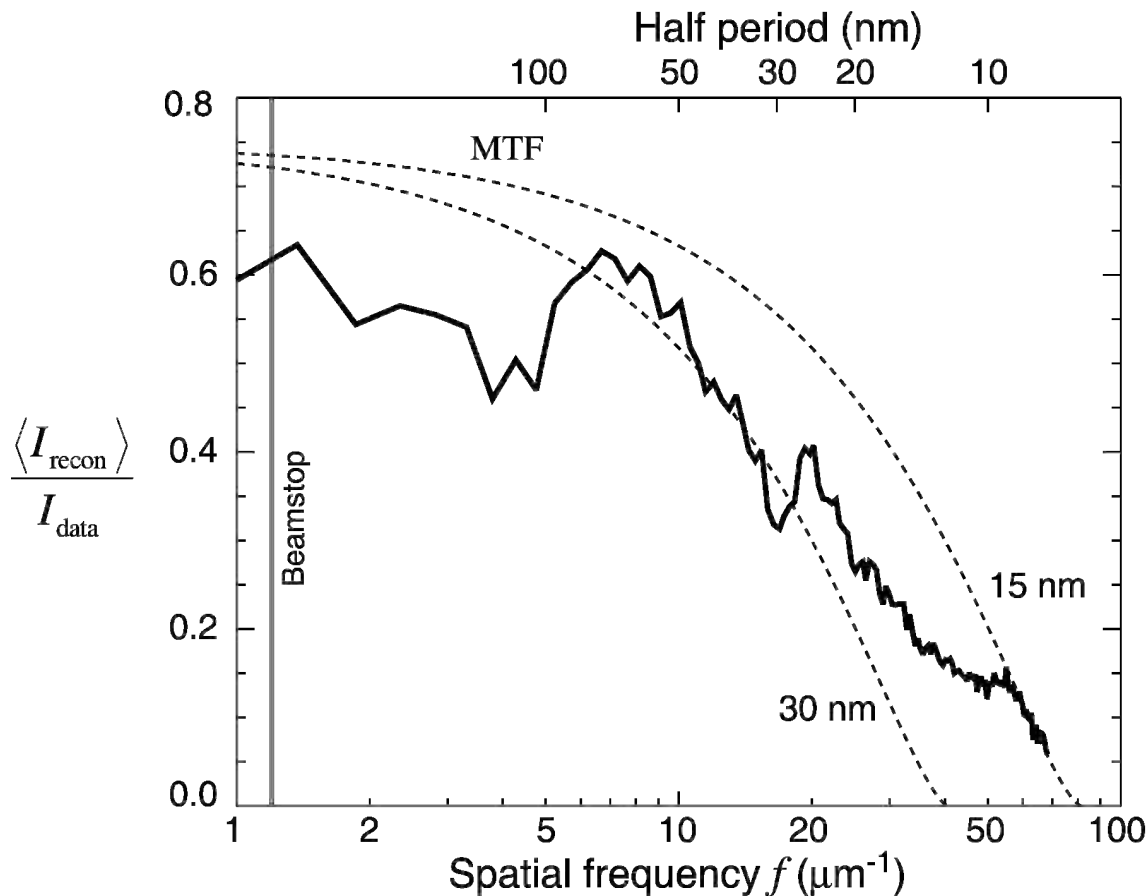
Diving deeper into reconstructions



(Erik Jacobsen, HMS Rhone, British Virgin Islands)

What is the resolution?

- We do not claim maximum angle of speckles as the resolution of the reconstructed image
- Instead, one can look at how much iterate preserves Fourier intensities at various spatial frequencies - like an MTF
- Elser and Thibault, Cornell



Problems we wish we didn't have...

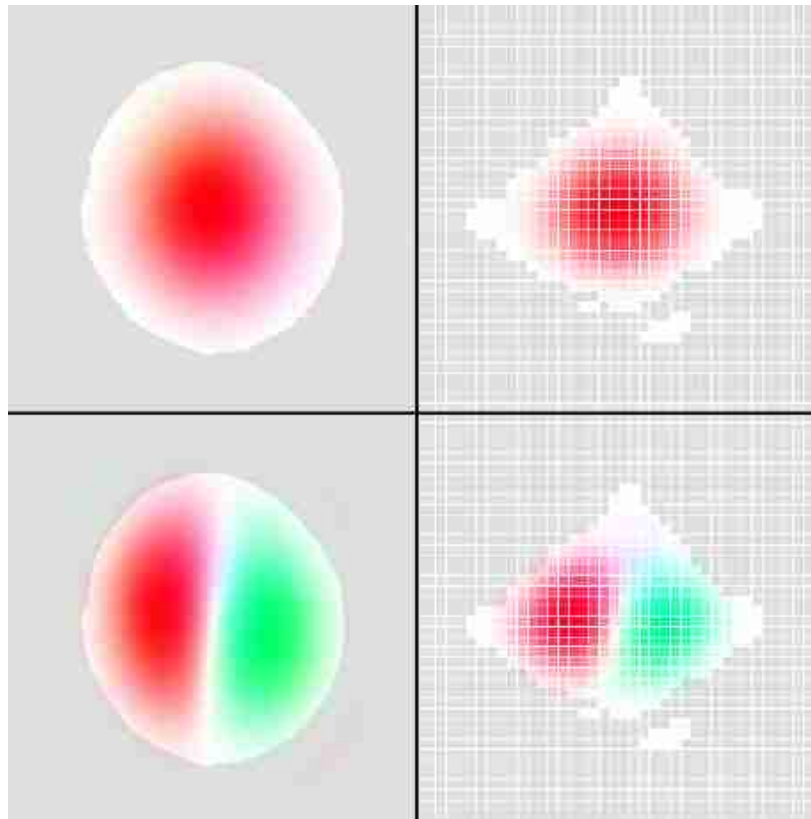


(The IGeSA beach)

Have my jellyfish! I'm not sellyfish... Ogden Nash

We miss those low frequencies...

Unconstrained modes in the reconstruction. See poster by Thibault et al.

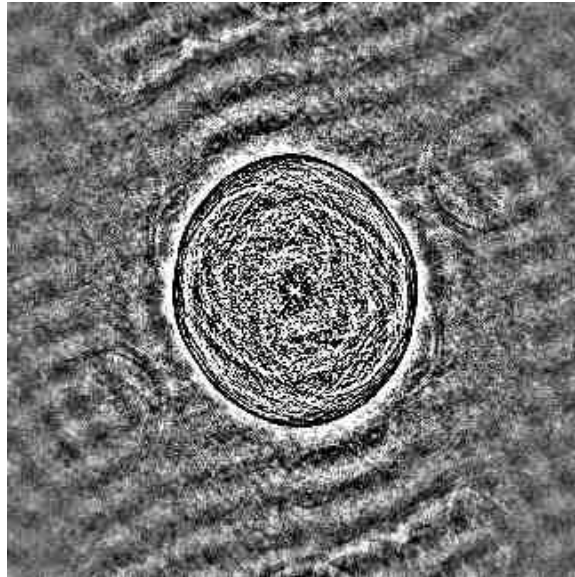


Real space

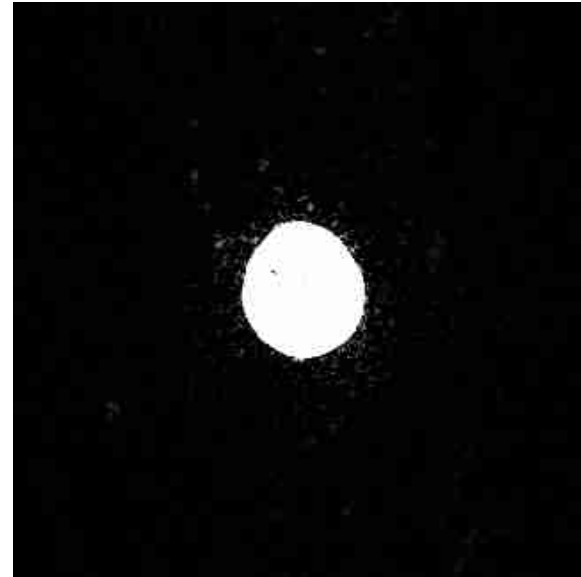
Fourier space

Is it really empty outside the support?

(Elser and Thibault, Cornell)



Real part of the
(Laplacian filtered)
autocorrelation of the
yeast cell data showing
evidence for the
presence of dirt around
the cell.



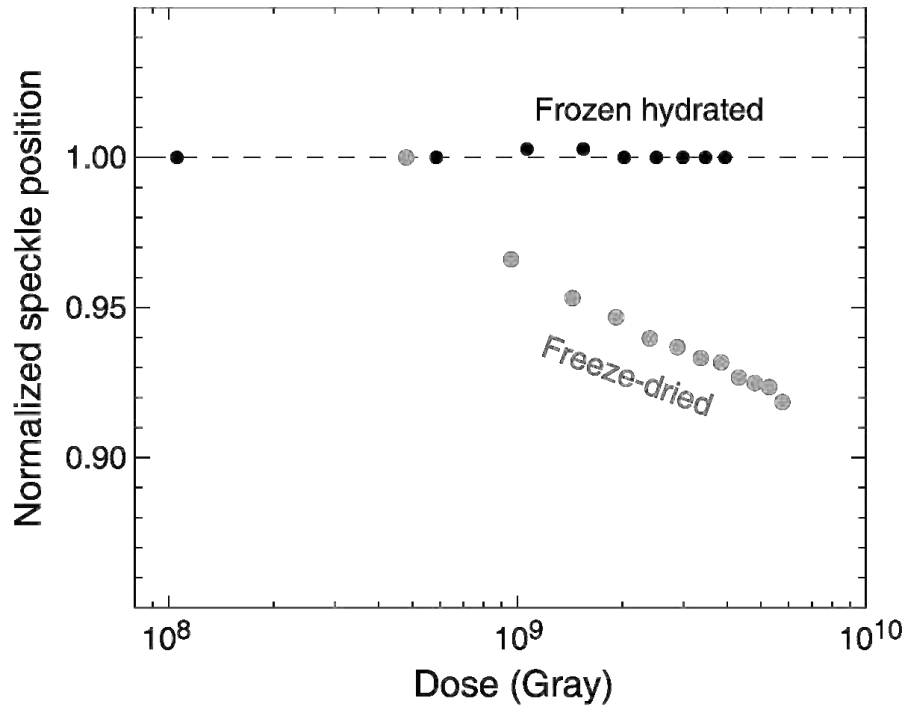
"Dirt" recovered around
the yeast cell. A
threshold-based support
has been added to the
fixed support of the cell.

Problems looming in the distance...

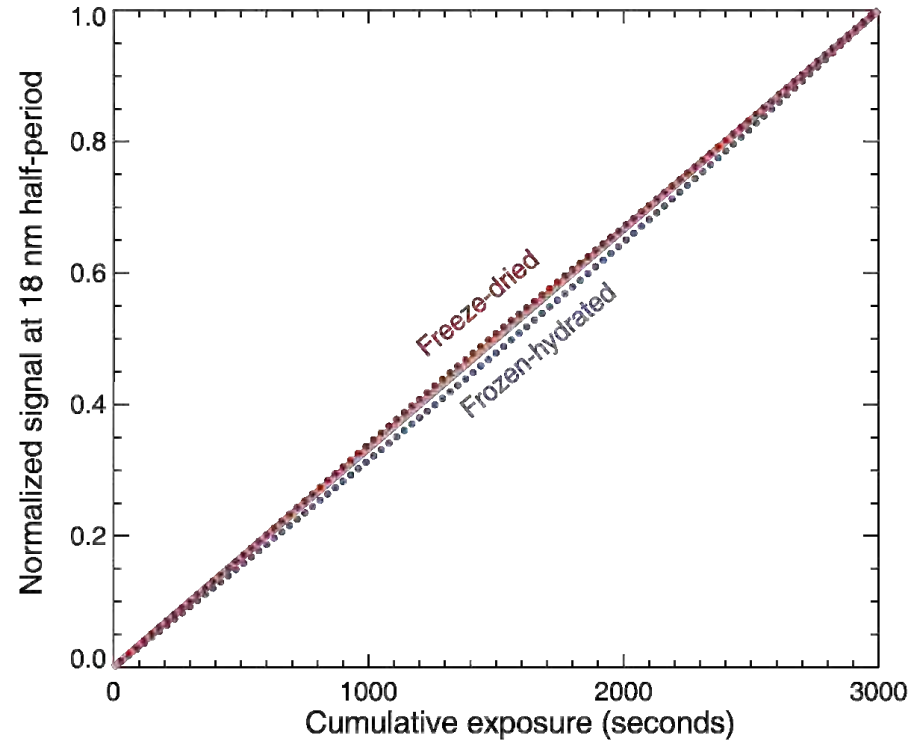


(Wreck Alley, British Virgin Islands)

Frozen hydrated?



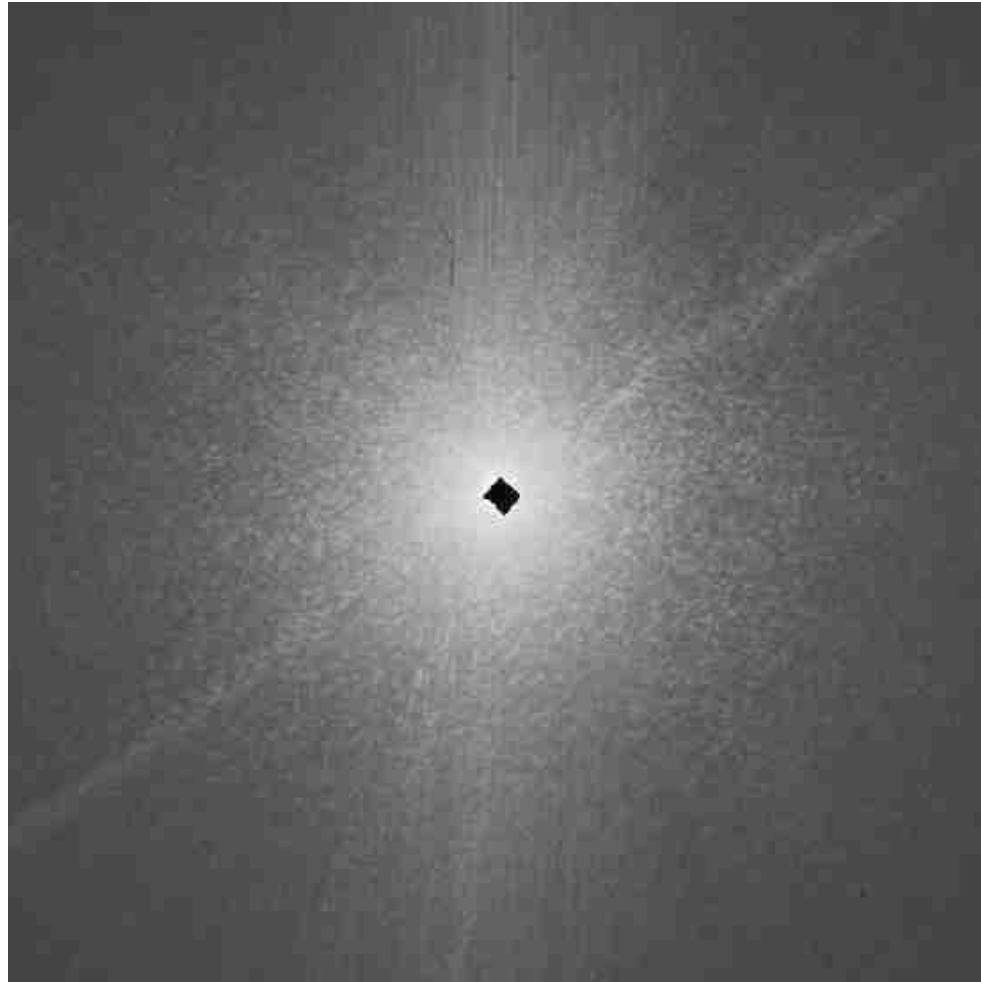
Frozen hydrated specimens don't shrink in the beam (freeze-dried specimens do)



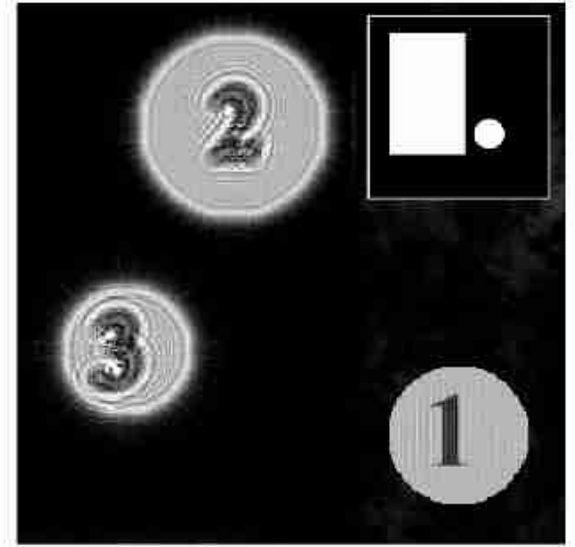
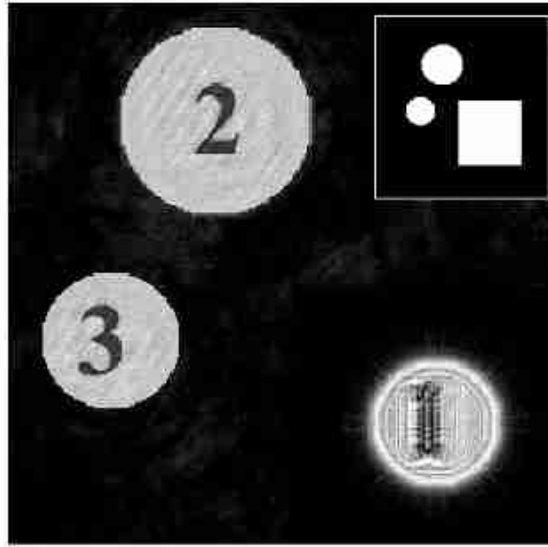
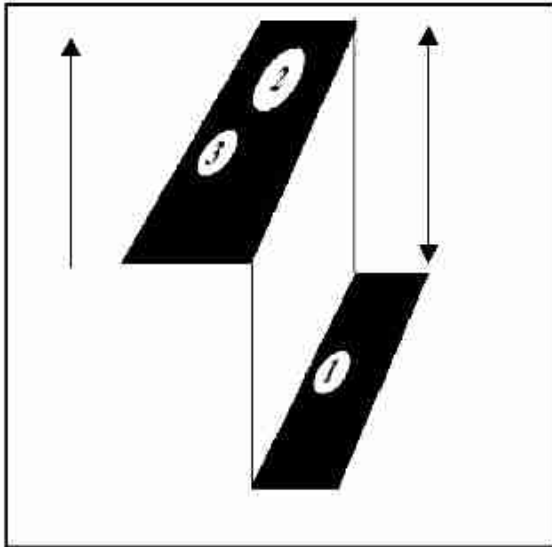
Scattering power is linear with dose thus far in both cases

Frozen hydrated!

Specimen preparation has been challenging... E. Lima and X. Huang, May 2005, at 520 eV.



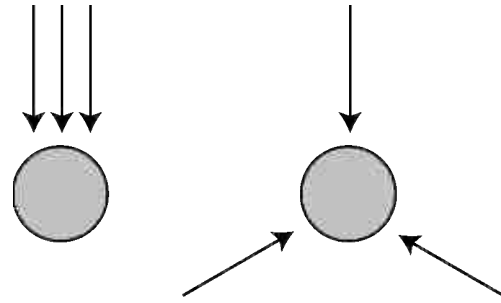
In 2D, what depth plane are we reconstructing?



Spence et al., Phil. Trans. R. Soc. London A **360**, 875 (2002)

3D reconstructions

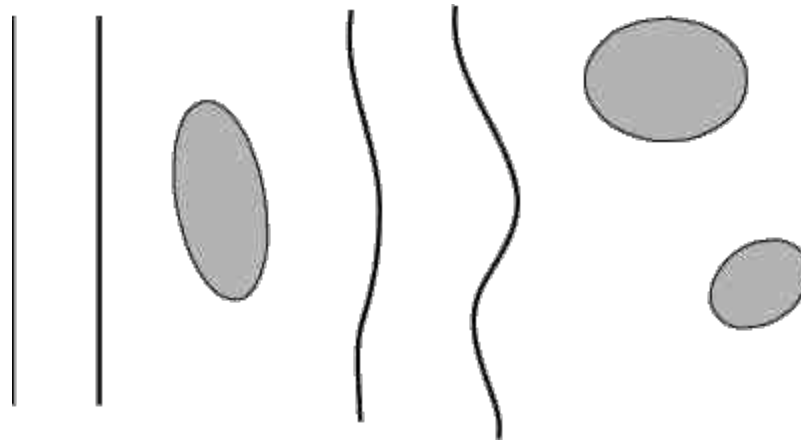
- Thanks to Chapman et al., we know it will work on our apparatus!
- Thanks to "dose fractionation", we know we don't need many more photons. Hegerl and Hoppe, *Z. Naturforschung* **31a**, 1717 (1976); McEwen et al., *Ultramic.* **60**, 357 (1995).



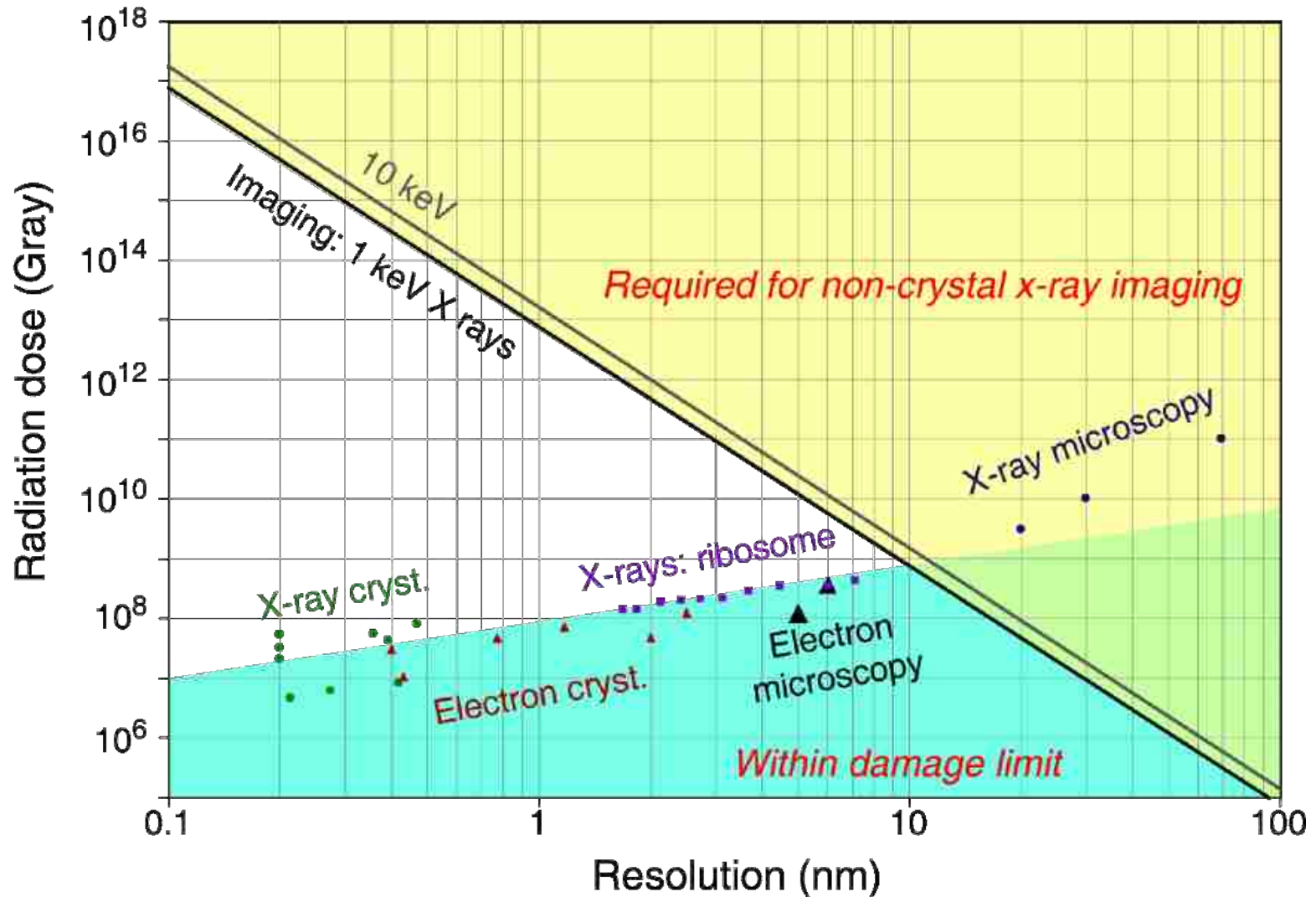
- We need to do more to automate our data taking, and work around CCD dynamic range limits (Tower of Hanoi?)
- Iterative phasing works better in 3D!
- A voxel is thinner than a pixel and thus less complex

Will we need to be Born again?

- Illumination of downstream planes can be corrupted by upstream objects
- Multislice calculations can be used to evaluate this problem; see poster by Thibault et al.
- Going to higher photon energies will help, though contrast then goes down



What's the limit for cells?



Thanks for your attention!



(HMS Rhone, British Virgin Islands)

Thanks for support from NIH, DoE, and NSF!